

Overview, Synthetic Biology

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Stanford Bioengineering
The BioBricks Foundation

9 July 2009

The U.S. National Academies
The Organization for Economic Cooperation and Development
The Royal Society

Overview, Synthetic Biology

What & why is synthetic biology?

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What and why is synthetic biology?

The recent and ongoing interest in 'synthetic biology' is being driven by at least four different groups: biologists, chemists, 're-writers' and engineers. Briefly, for biologists, the ability to design and construct synthetic biological systems provides a direct and compelling method for testing our current understanding of natural biological systems^{4,15}; disagreements between expected and observed system behaviour can serve to highlight the science that is worth doing. For chemists, biology is chemistry, and thus synthetic biology is an extension of synthetic chemistry; the ability to create novel molecules and molecular systems allows the development of useful diagnostic assays and drugs, expansion of genetically encoded functions, study of the origins of life, and so on¹⁶. For 're-writers', the designs of natural biological systems may not be optimized for human intentions (for example, scientific understanding, health and medicine); synthetic biology provides an opportunity to test the hypothesis that the genomes encoding natural biological systems can be 're-written', producing engineered surrogates that might usefully supplant some natural biological systems¹¹. Finally, for engineers, biology is a technology; building upon past work in genetic engineering, synthetic biology seeks to combine a broad expansion of biotechnology applications with—as the focus of this article—an emphasis on the development of foundational technologies that make the design and construction of engineered biological systems easier.

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- *Natural Science*
- *Synthetic Science*
- *Re-Writers*
- *Engineers*

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- *Natural Science*
- *Synthetic Science*
- *Re-Writers*
- *Engineers*
- *Humanity!*

The nature of synthetic biology

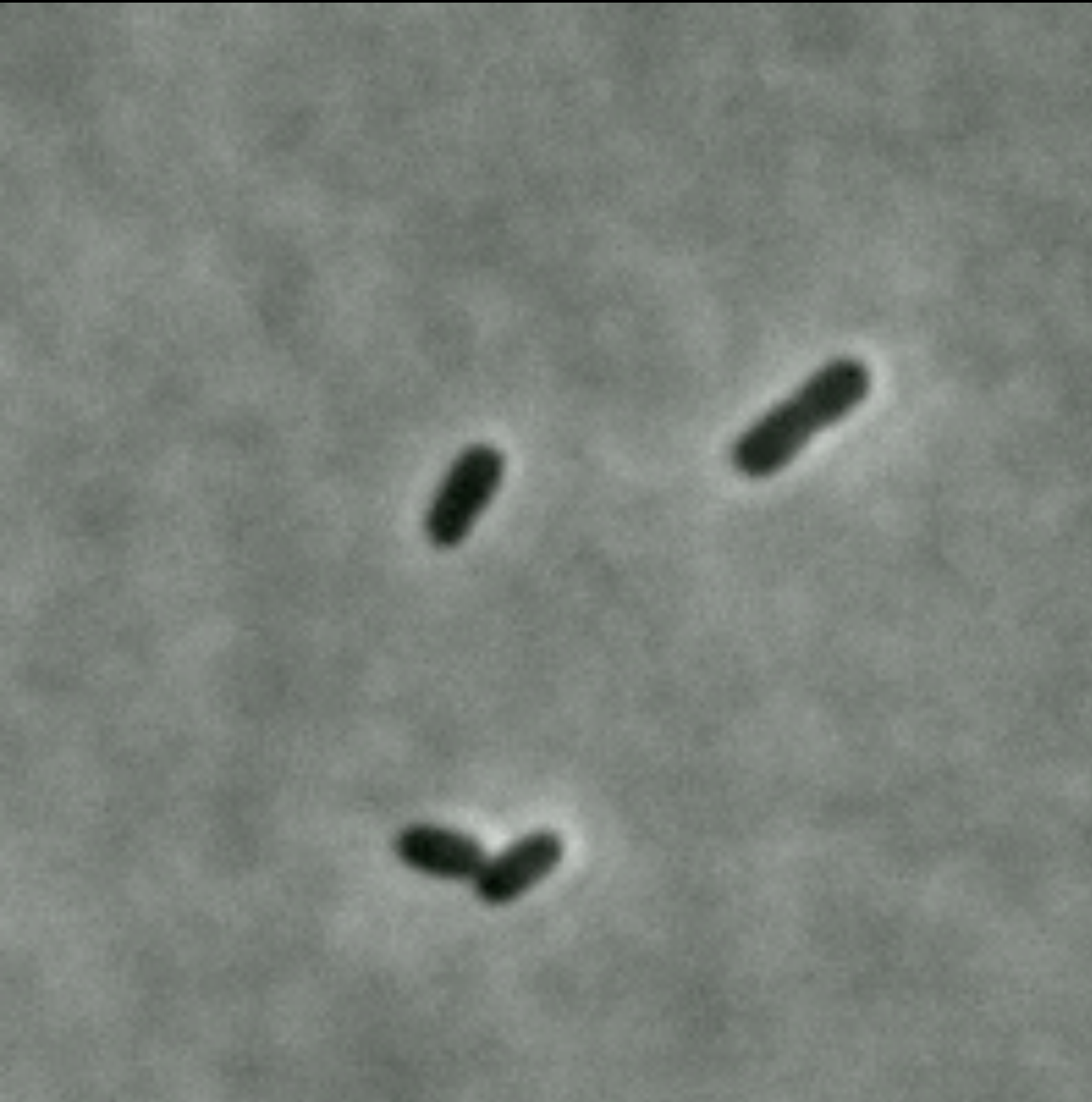
The nature of synthetic biology



<http://www.cliquee.net/wp-content/uploads/2008/01/disassembly.jpg>

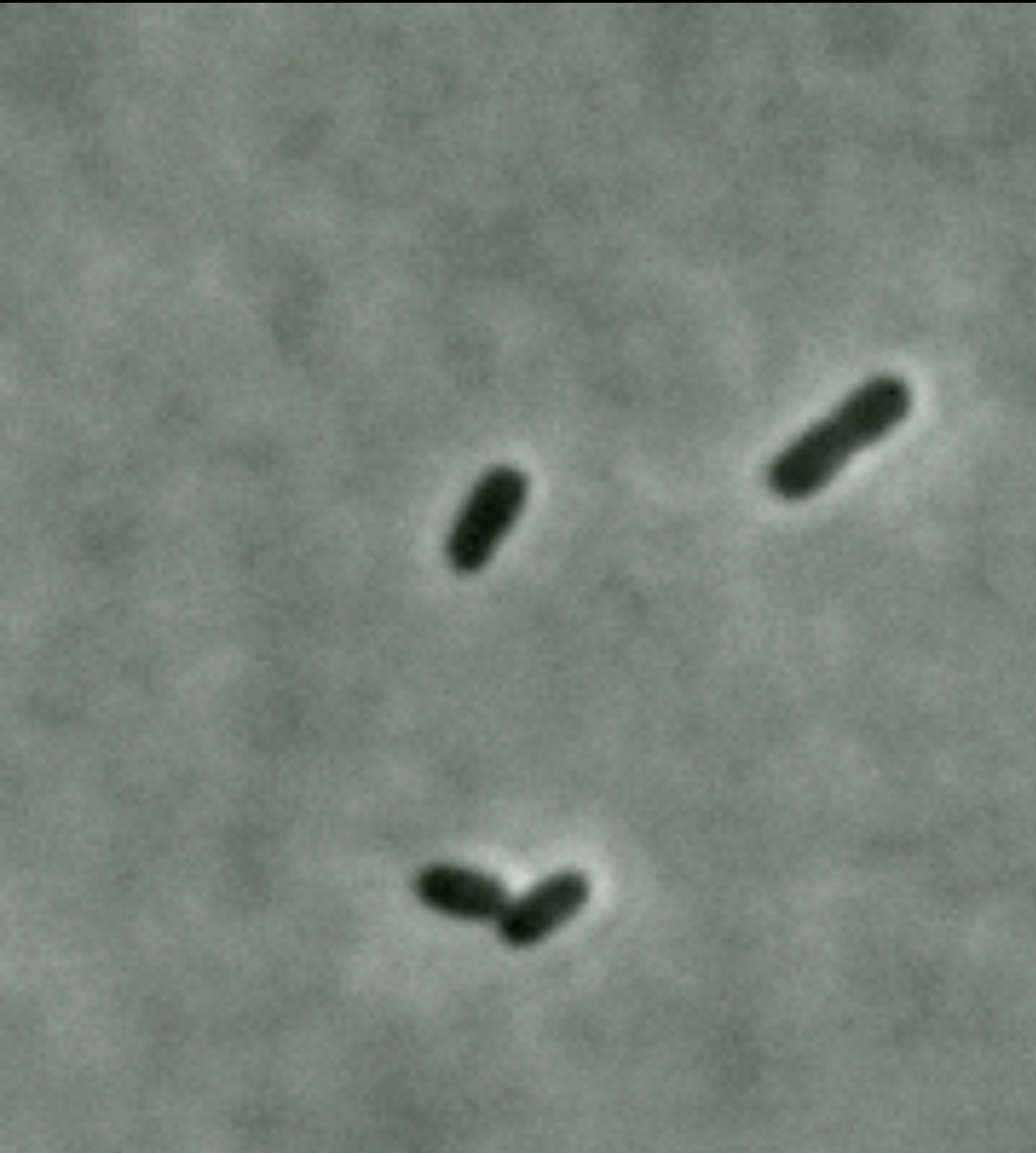
The **nature** of synthetic biology

The **nature** of synthetic biology



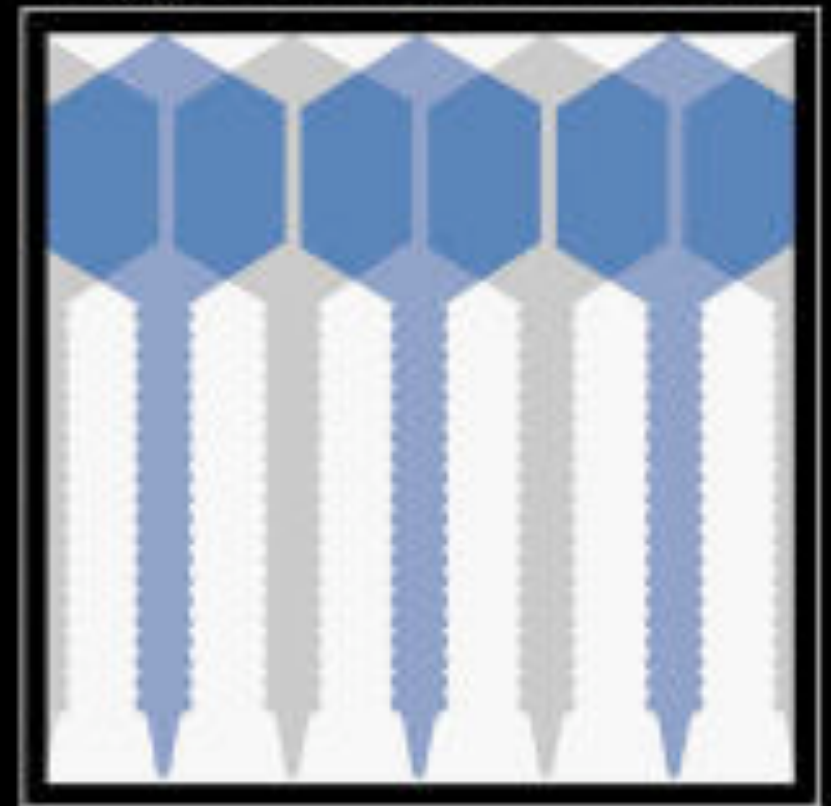
St-Pierre & Endy, PNAS USA (2008)

The **nature** of synthetic biology



St-Pierre & Endy, PNAS USA (2008)

A
GENETIC
SWITCH
Third Edition
Phage Lambda Revisited



MARK PTASHNE

THE DECISION

Having described the two developmental pathways available to an infecting λ phage, we now must ask: what determines which pathway is taken? What factors drive the system toward lysis or lysogeny?

We do not have a complete understanding of these matters but we can construct a plausible scenario. Briefly put, the “decision” is effected by a single protein—CII.

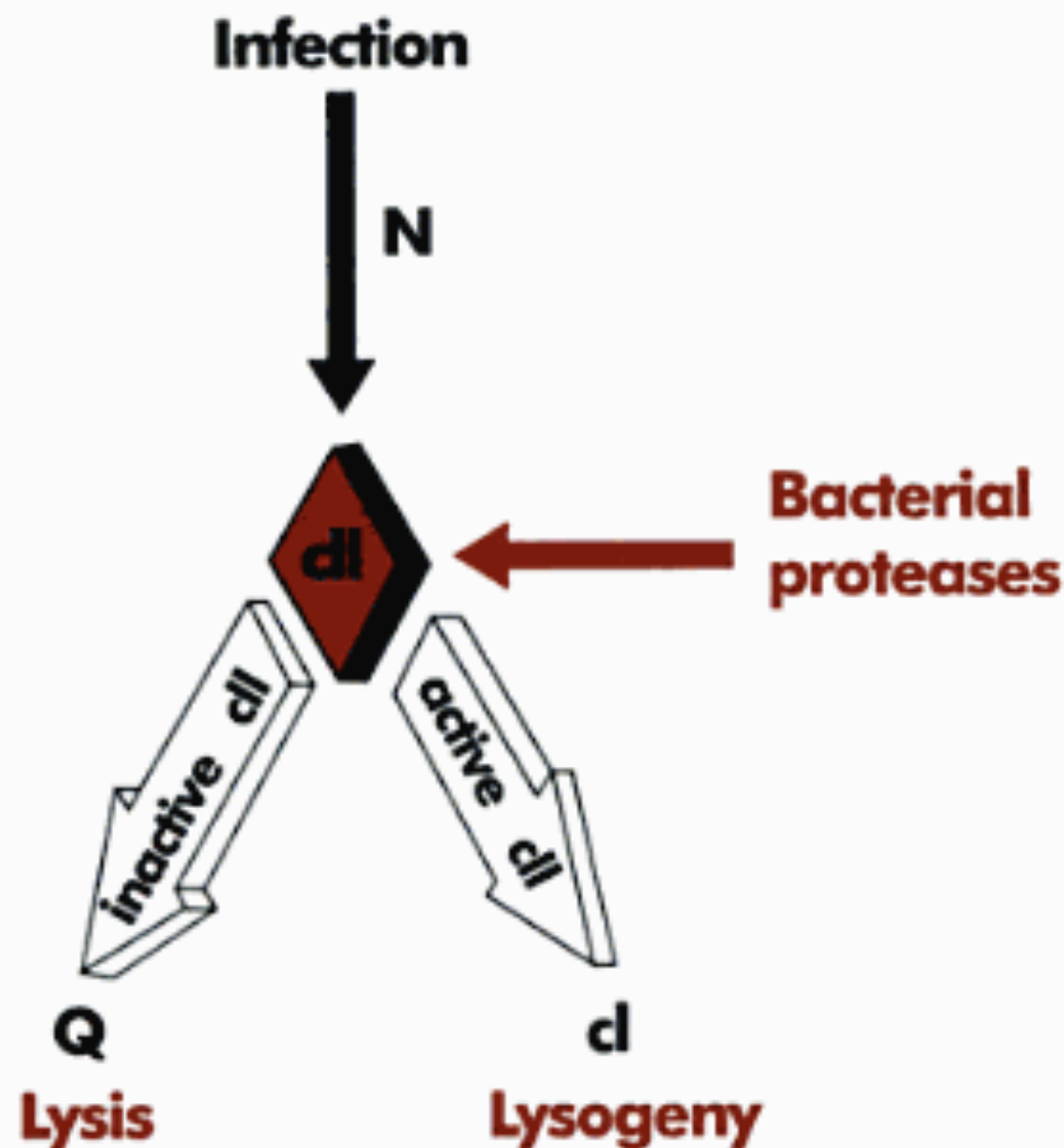


Figure 3.10. The lysis-lysogeny decision. Host proteases regulate the level of activity of CII protein. Although CIII protein is not shown here, the host factors may exert their effects by working on CIII, which protects CII. It is likely that other host proteins regulate translation of the CII mRNA as well.

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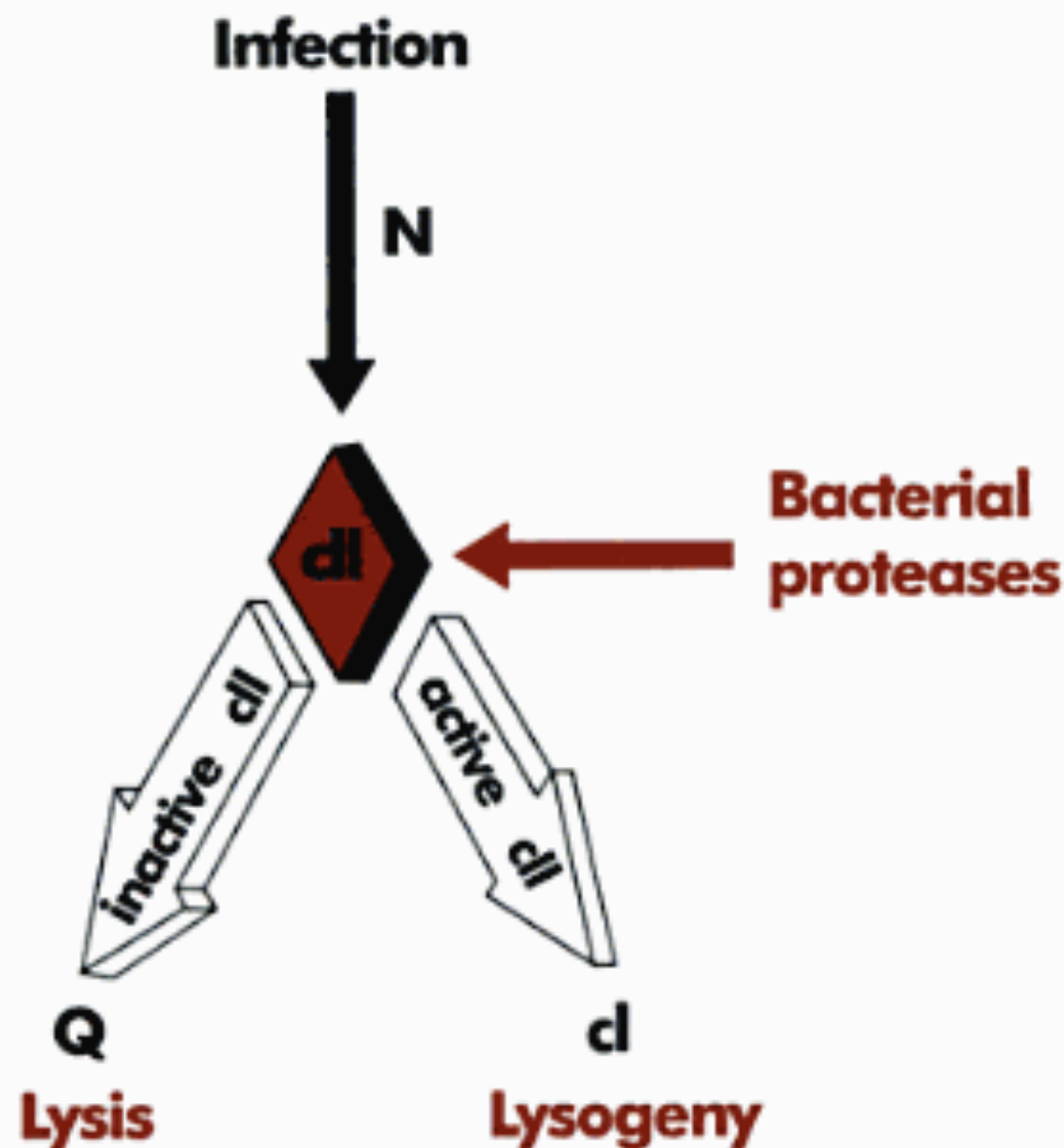
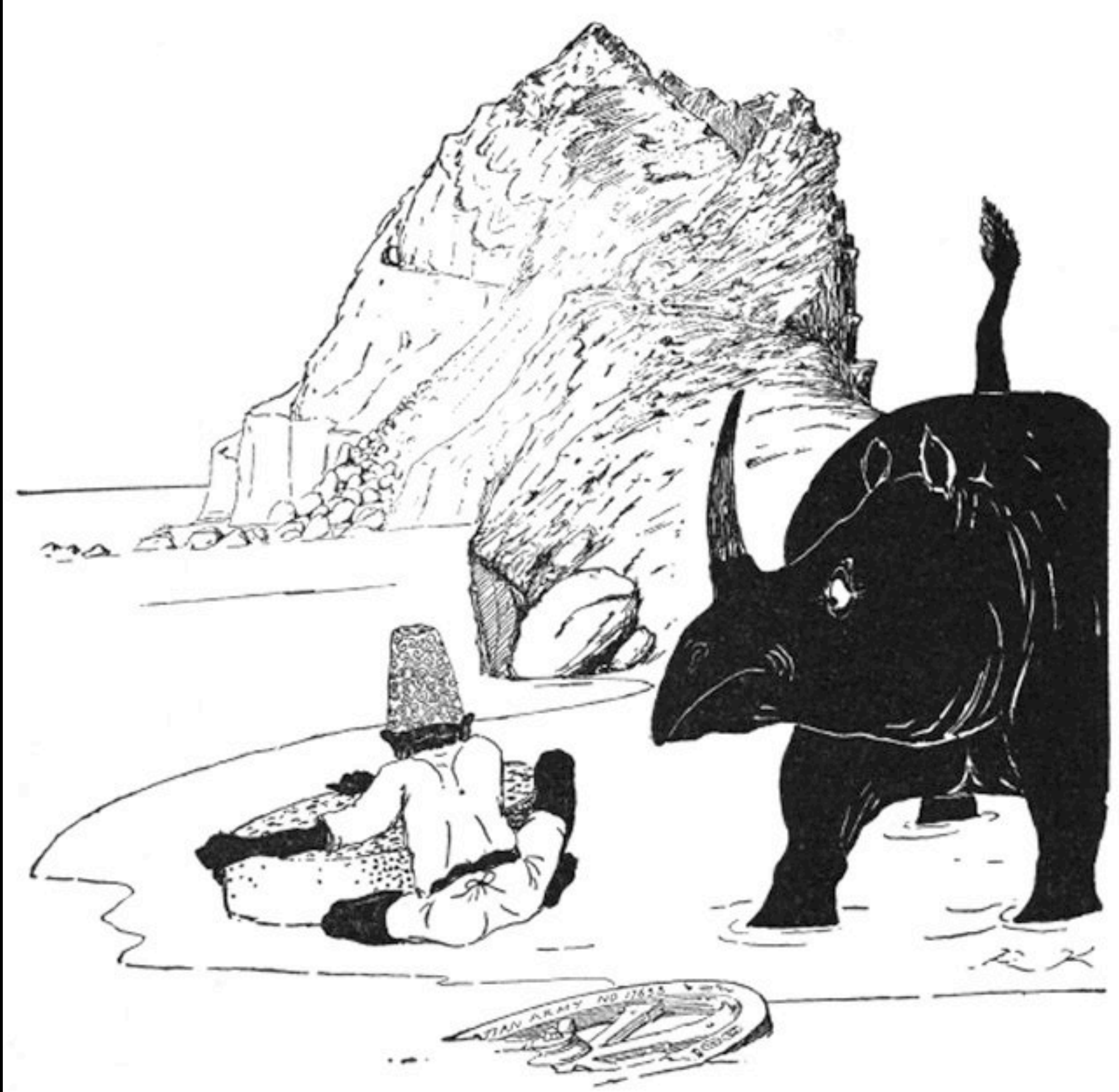


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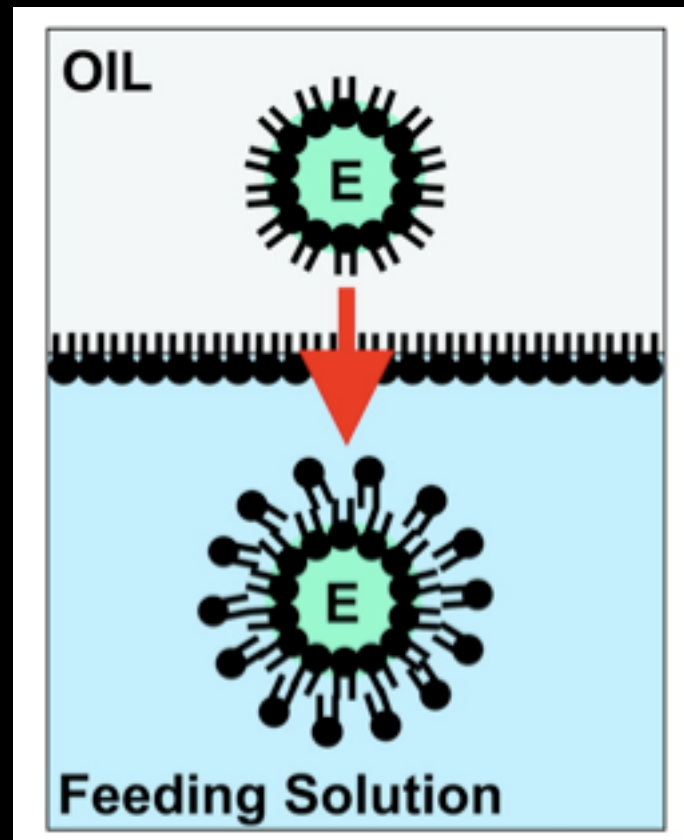


Understanding **nature** via synthesis

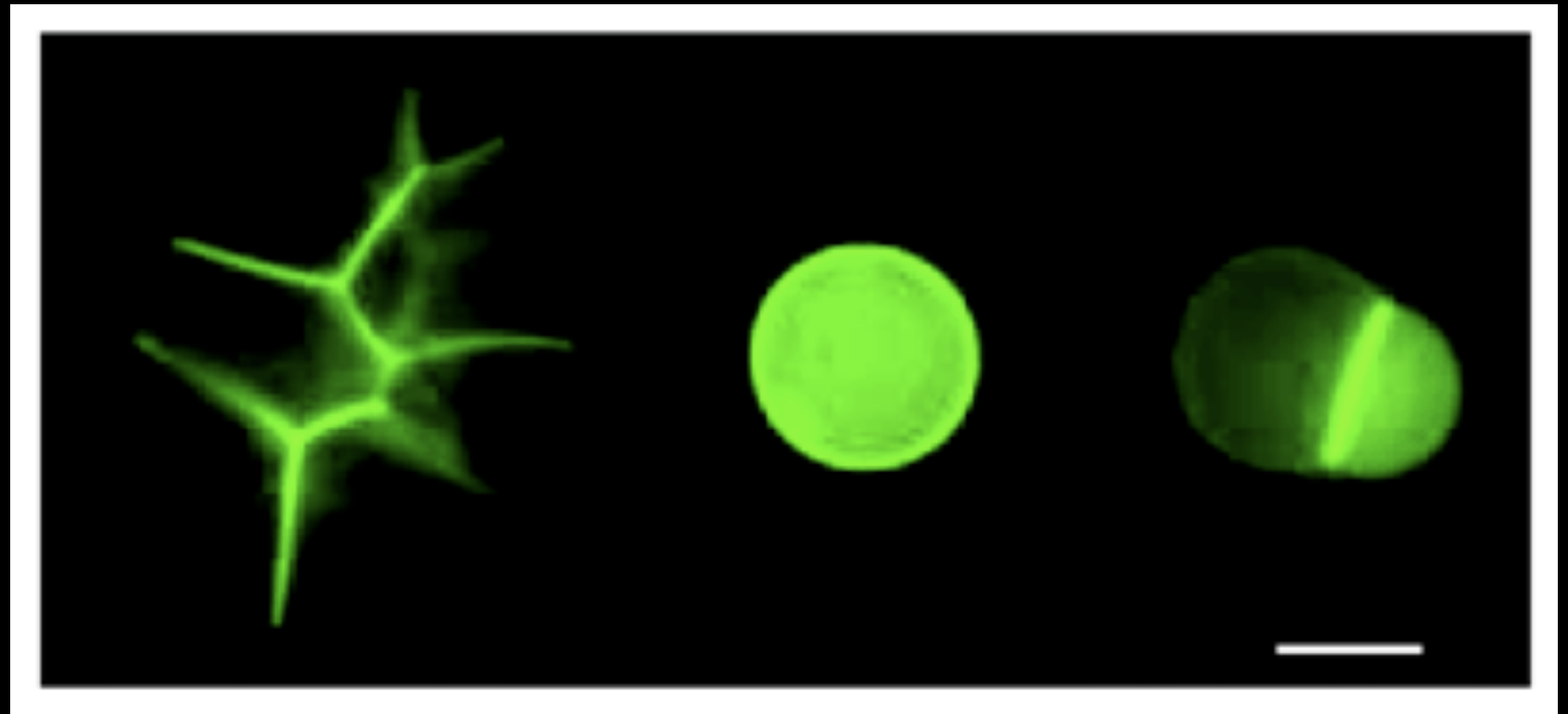
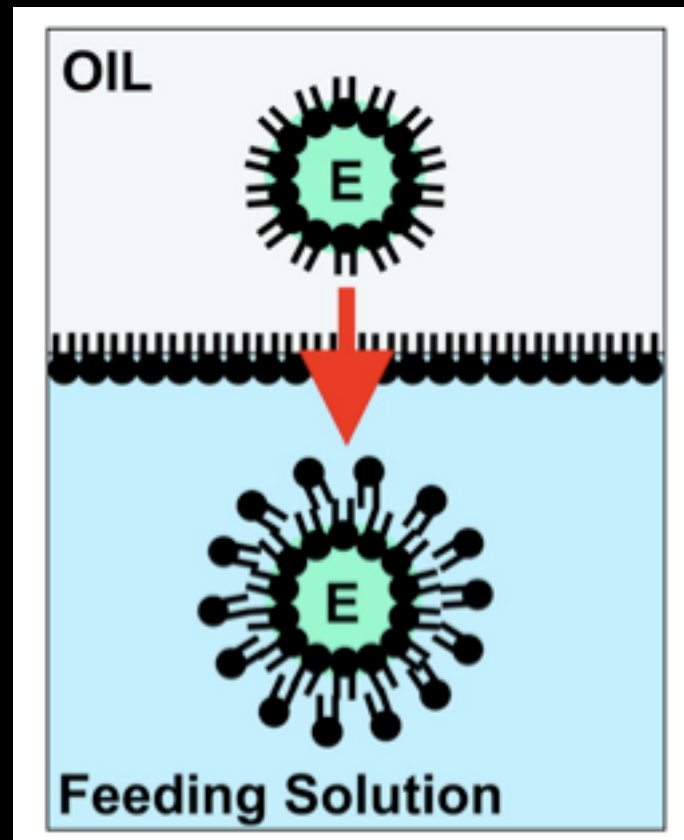
Understanding **nature** via synthesis

The **synthesis** of synthetic biology

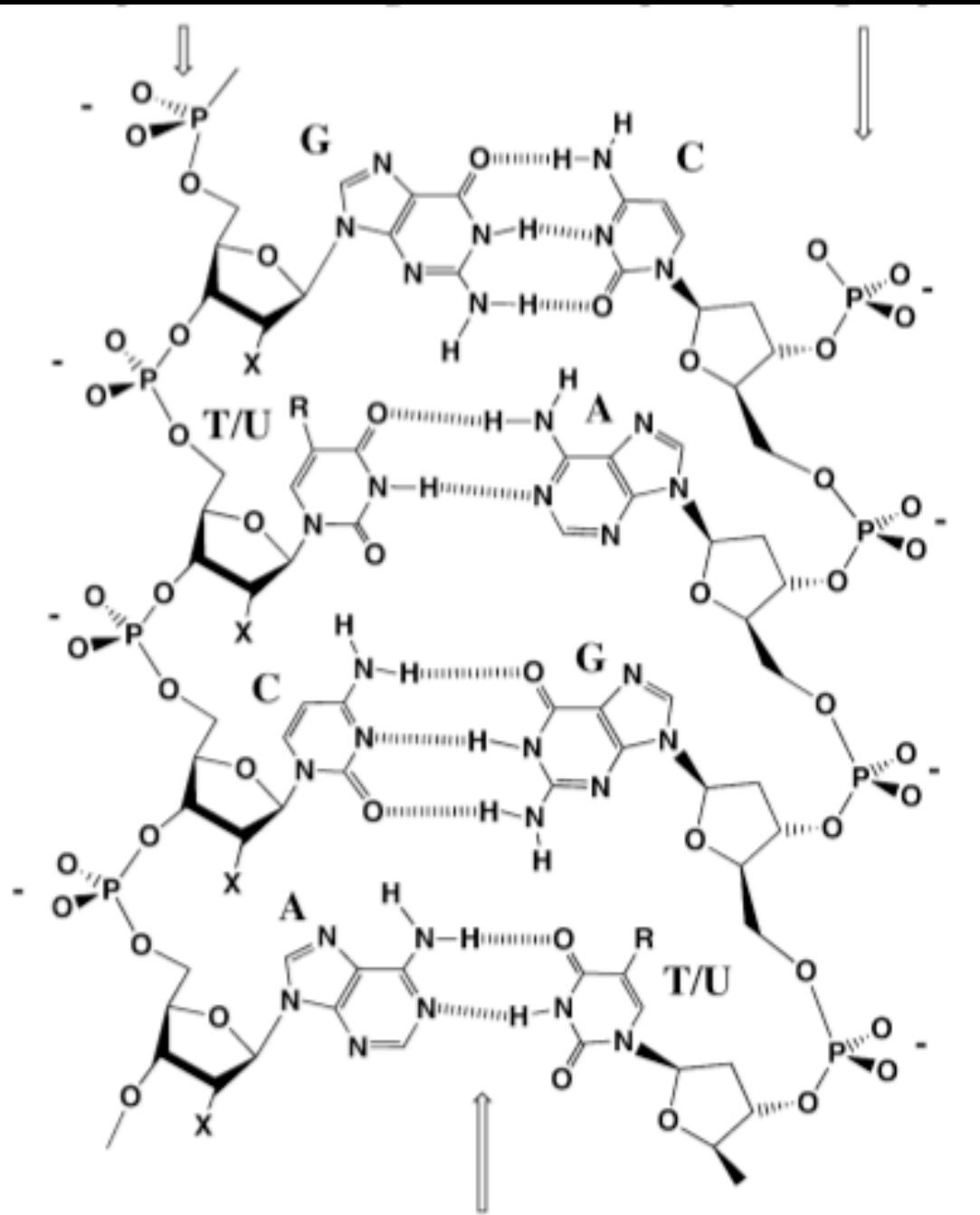
The **synthesis** of synthetic biology



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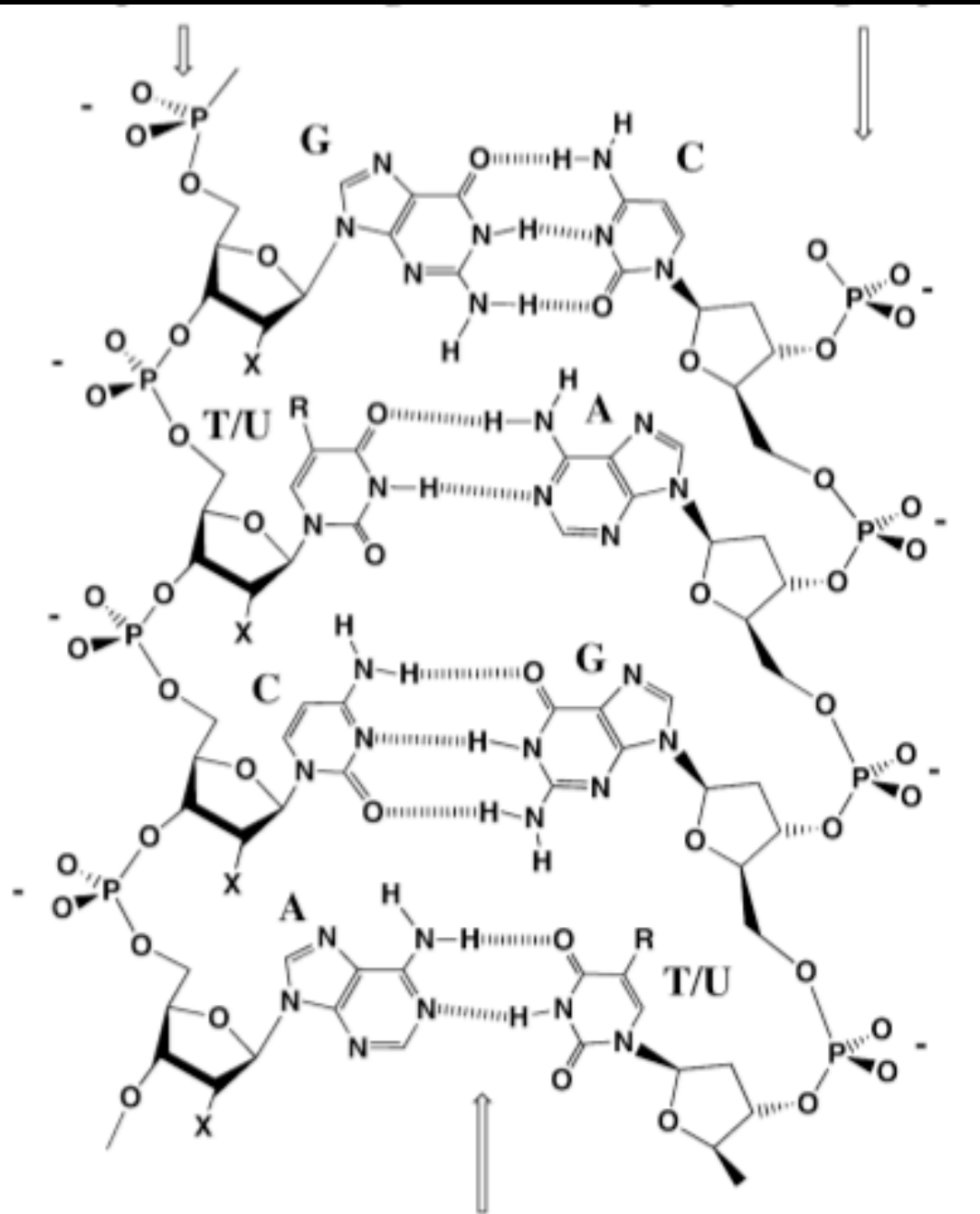


forces interstrand contact here, as far from the backbone as possible

FIGURE 14. Backbone charges force interstrand contacts in a DNA duplex to the Watson–Crick edge of the heterocycles, hinder folding, and dominate physical behavior, allowing DNA/RNA to mutate and evolve.

The **synthesis** of synthetic biology

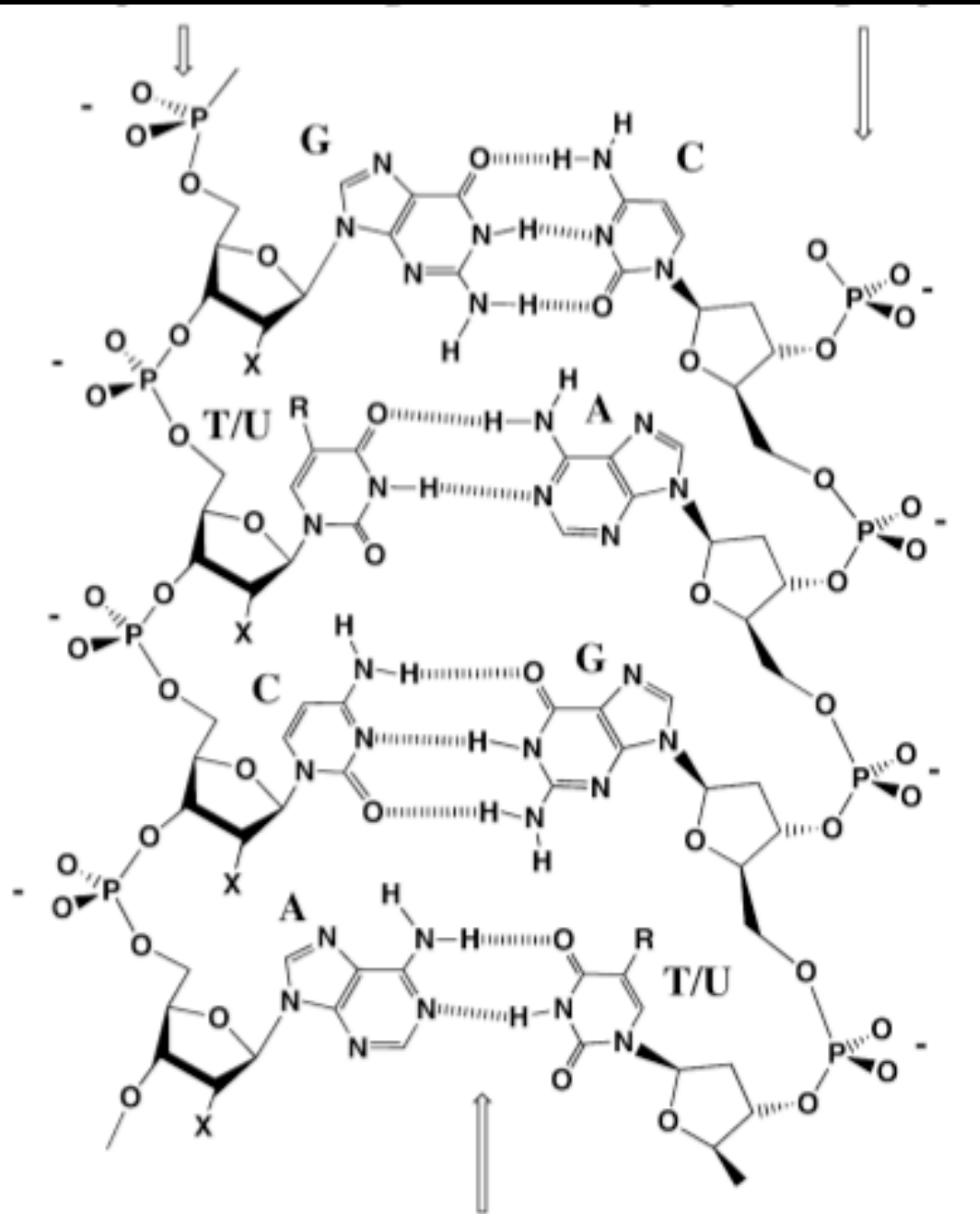
- Why repeated negative charges?



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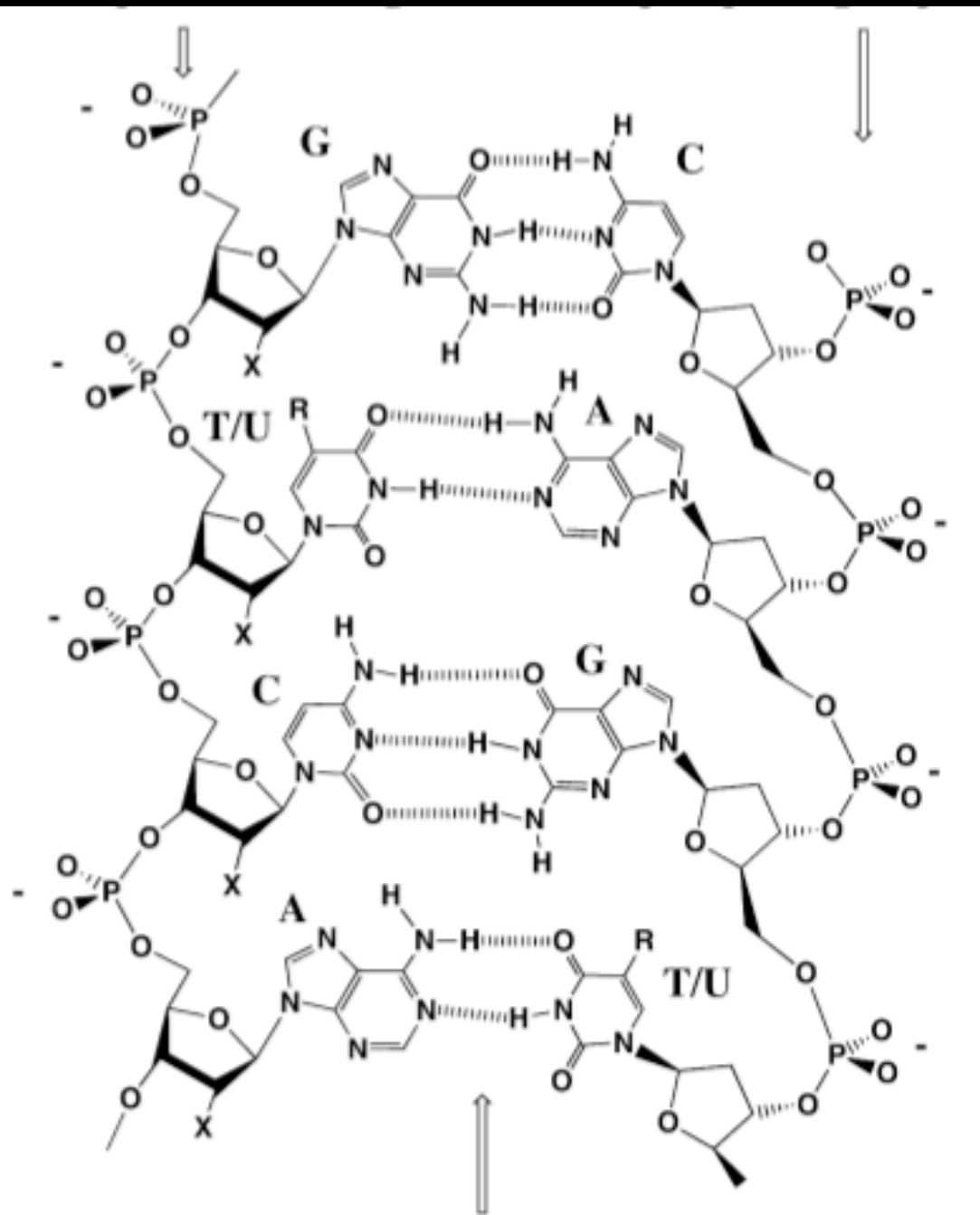


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- Why repeated negative charges?
- Replace anionic phosphodiester linkers with uncharged dimethylene sulfones
- Discover phosphate backbone hinders folding, enables base pairing, and allows for mutations

The **liberation** of synthetic biology

The **liberation** of synthetic biology



The **liberation** of synthetic biology

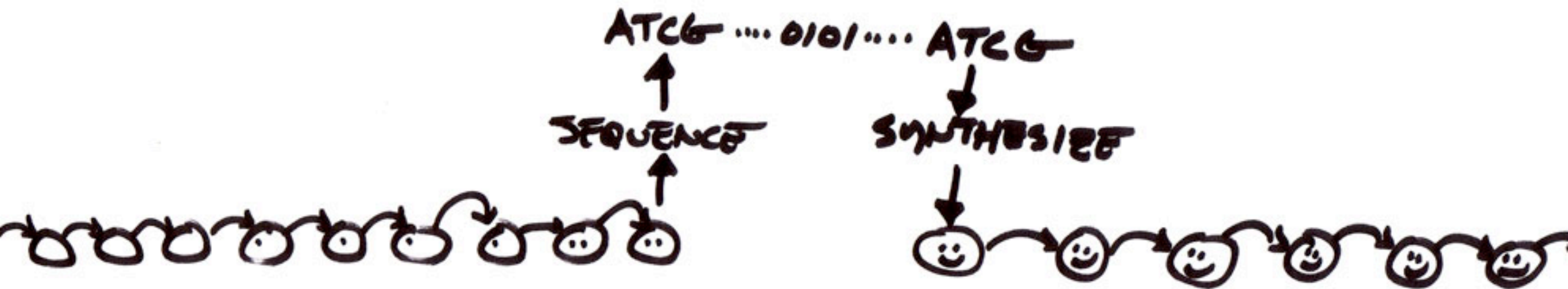


Evolution = Tyranny
(mutation without representation)

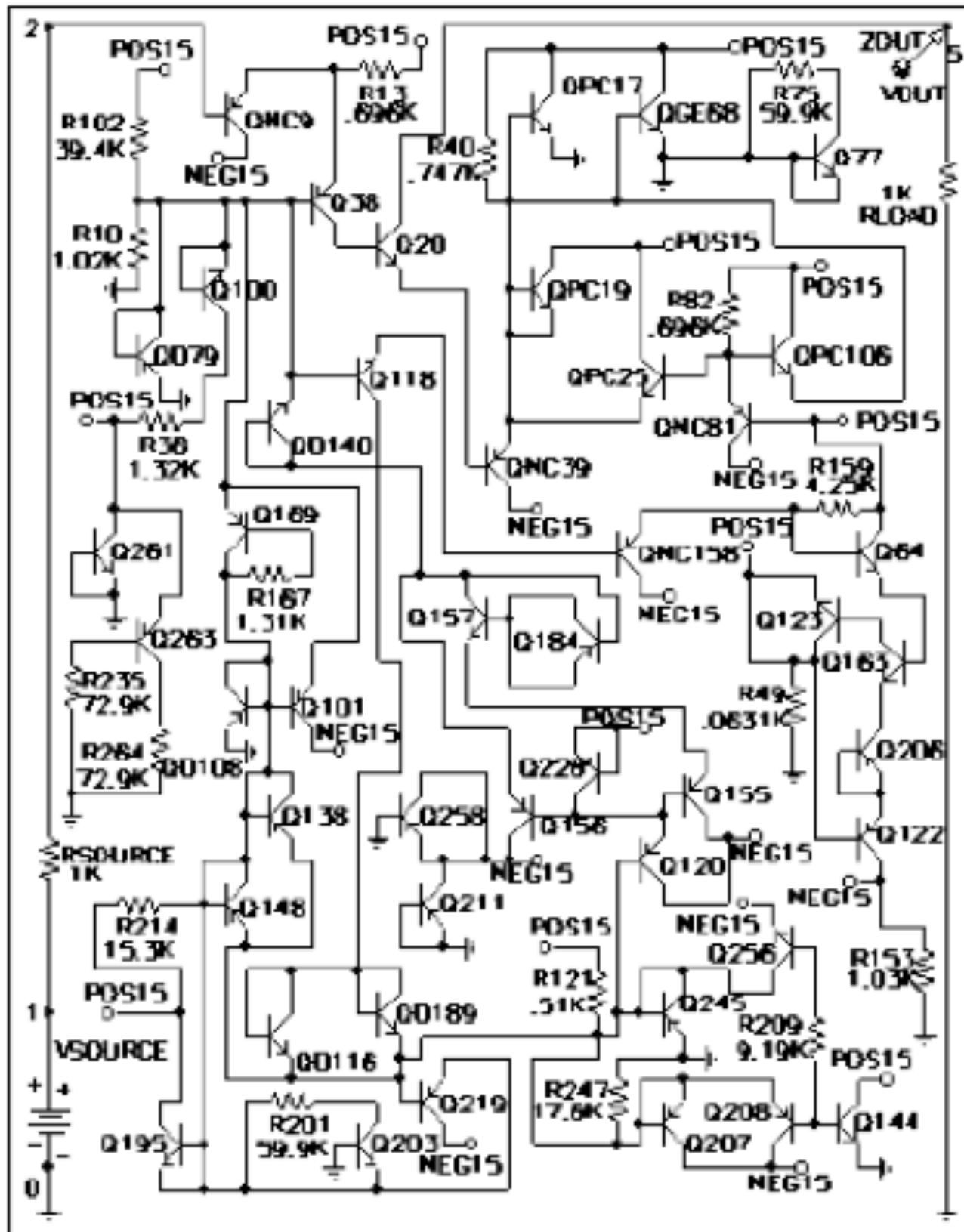
The liberation of synthetic biology



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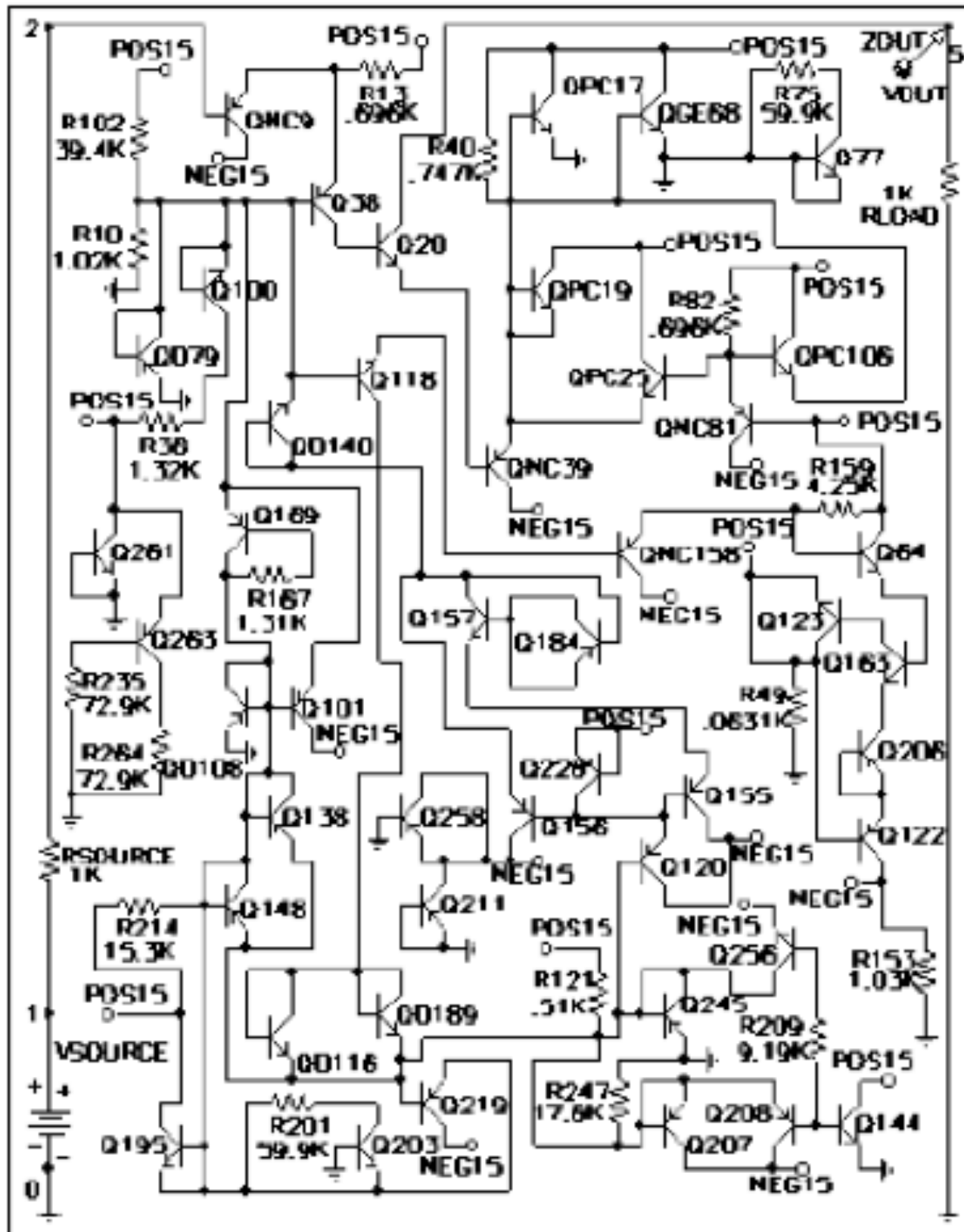


The liberation of synthetic biology



J.R. Koza et al.
Automated Synthesis of Computational
Circuits using Genetic Programming,
1997 IEEE International Conference on
Evolutionary Computation

The liberation of synthetic biology



1. What does it do?
2. How does it work?
3. Why this design?

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Automated Synthesis of Computational
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The liberation of synthetic biology

The liberation of synthetic biology

-----2.8----->
acgcaaaggaggcgacatggcagggttacggcgctaaaggaatccgaaa
<--3-RBS--><-----3-----

The liberation of synthetic biology

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acgcaaaggaggcgacatggcaggttacggcgctaaaggaatccgaaa

<--3-RBS--><-----3-----

acgcaaGgggagAcgacaCggcaggttacggcgctaaaggatcggccgcaaaggagggcgacatggcaggttacggcgctaaa

-----2.8-----><D28R | D29L><--3RBS-----><-----3-----

The liberation of synthetic biology

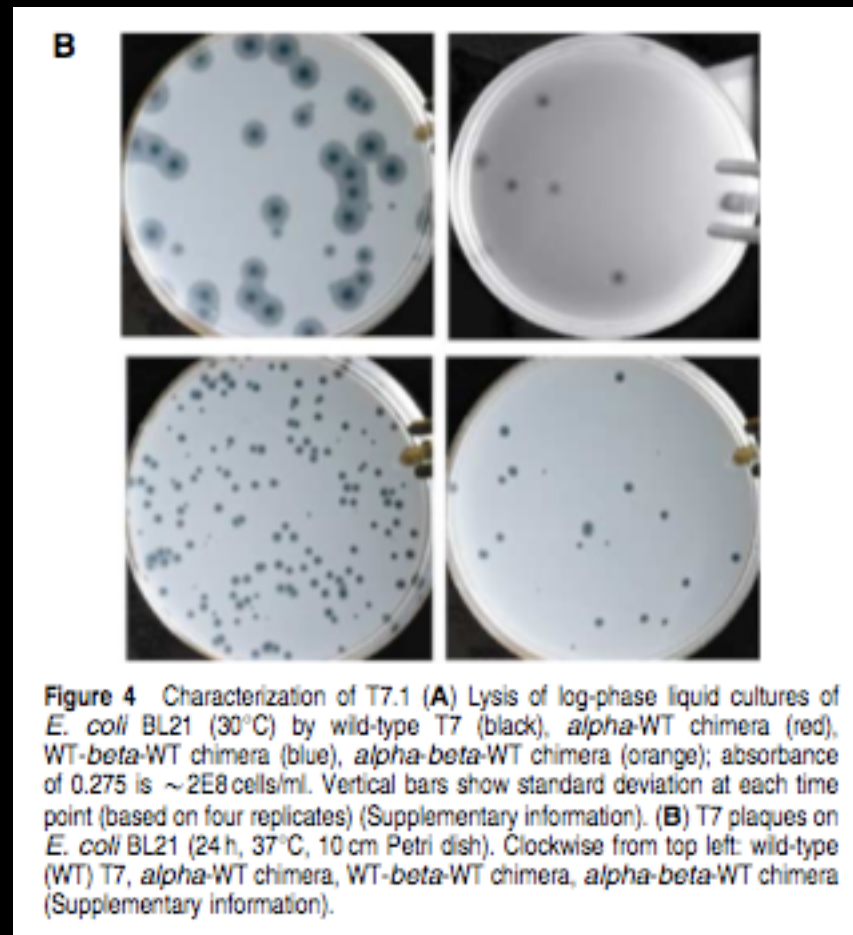
-----2.8----->

acgcaaaggaggcgacatggcaggttacggcgctaaaggaatccgaaa

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acgcaaGgggagAcgacaCggcaggttacggcgctaaaggatcggccgcaaaggaggaggcgacatggcaggttacggcgctaaa

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The engineering of synthetic biology

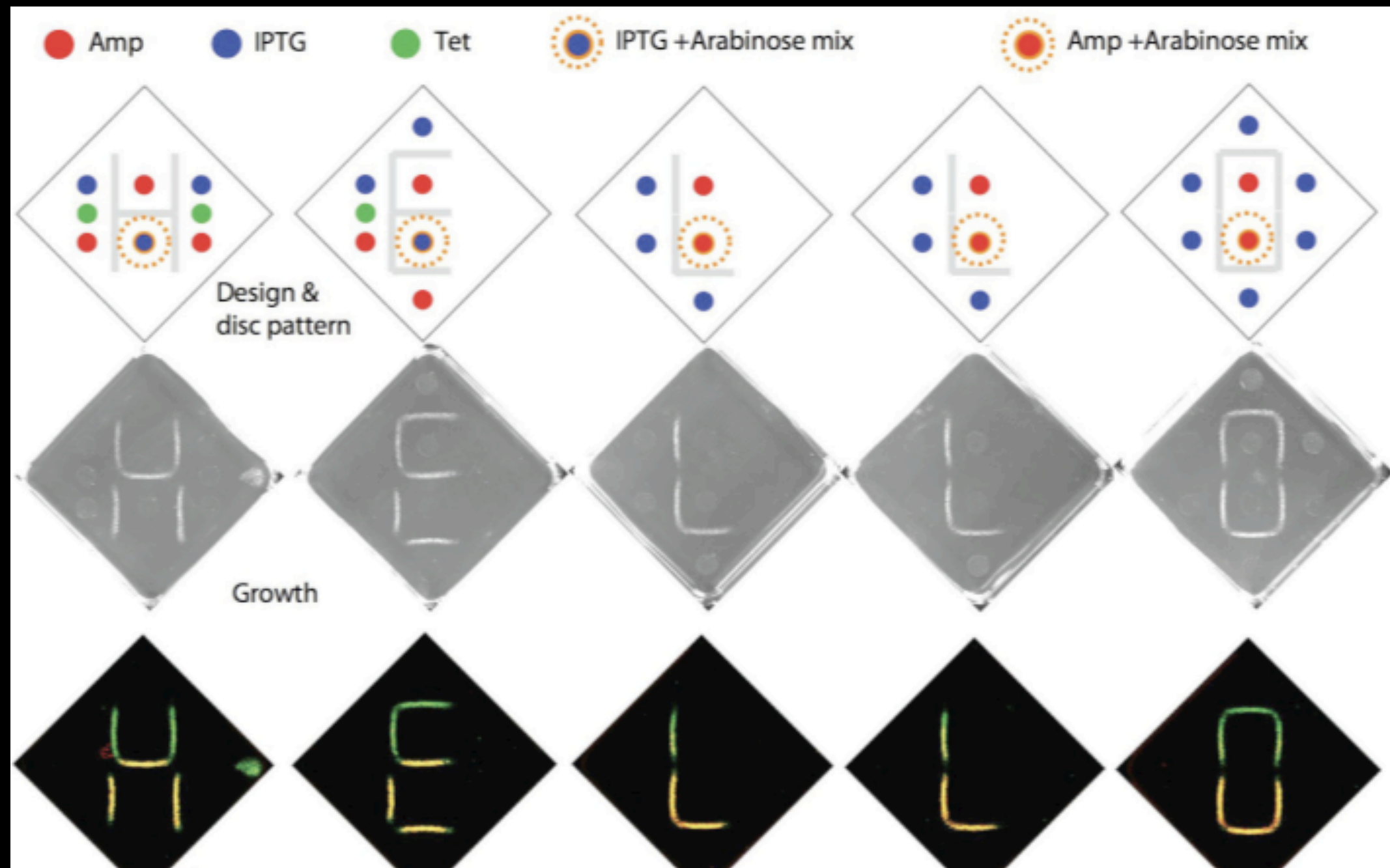
The engineering of synthetic biology



summer vocation:

"I Was a Teenage Genetic Engineer"

The engineering of synthetic biology



Morphogen-defined patterning of *Escherichia coli* enabled by an externally tunable band-pass filter

Journal of Biological Engineering 2009, **3**:10 doi:10.1186/1754-1611-3-10

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Richard A Heins (rheins2@jhu.edu)

Marc Ostermeier (oster@jhu.edu)

The engineering of synthetic biology

The **engineering** of synthetic biology

Teach me how to...

The engineering of synthetic biology

Teach me how to...

Design and build living organisms that behave as expected.

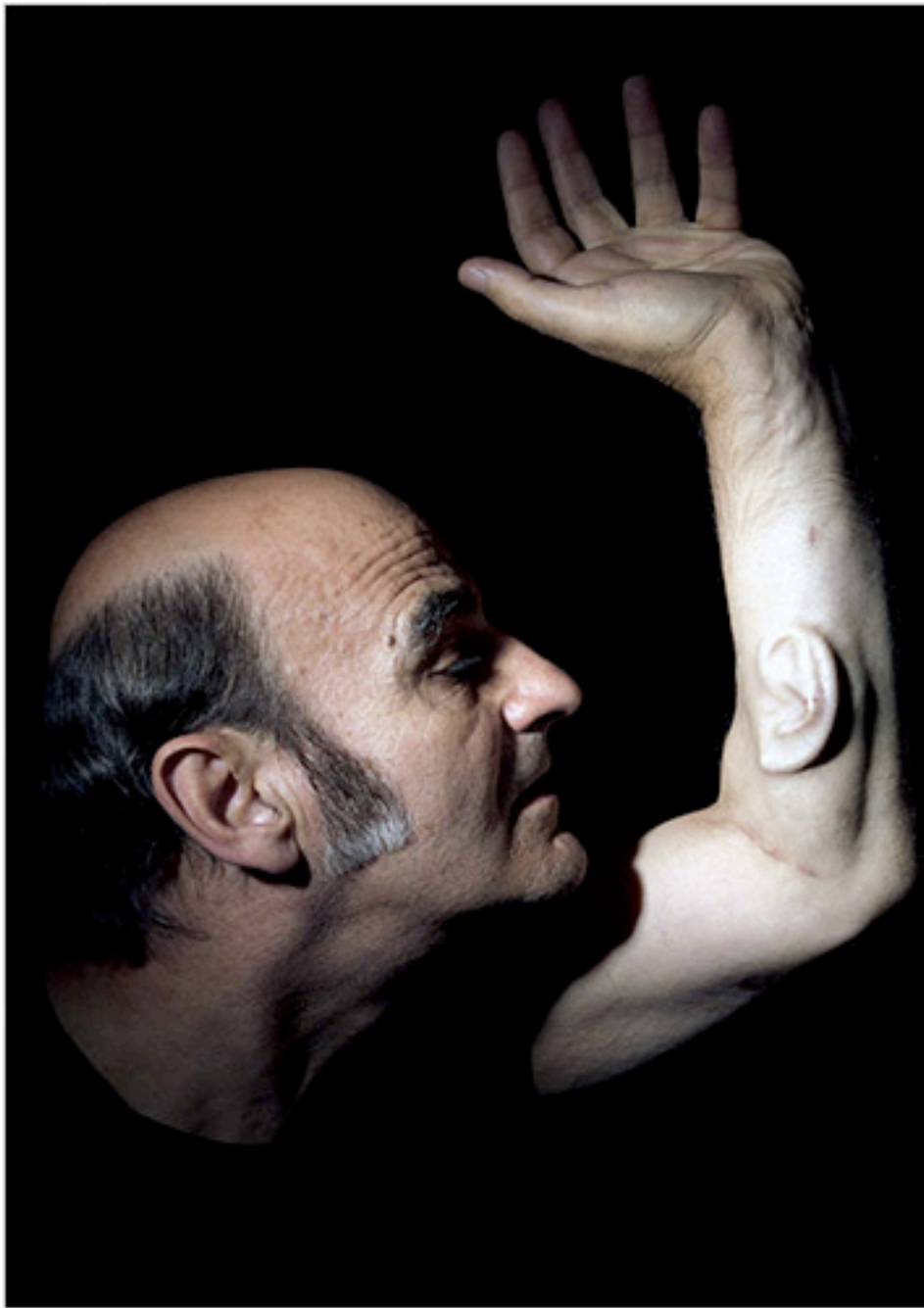
Debug existing or write new genetic programs to do my bidding.

The **humanity** of synthetic biology

The **humanity** of synthetic biology

The New York Times

April 14, 2009

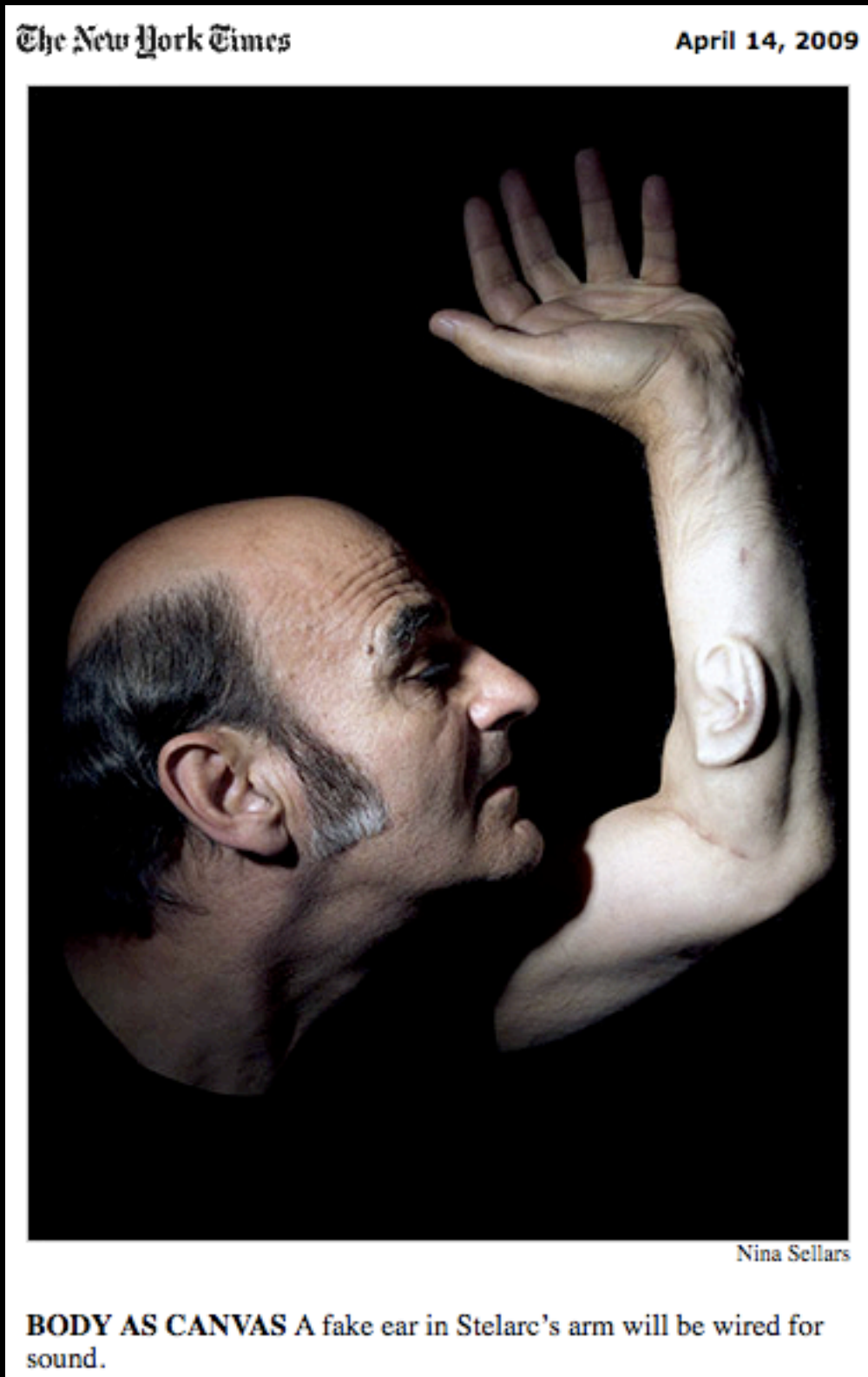


Nina Sellars

BODY AS CANVAS A fake ear in Stelarc's arm will be wired for sound.

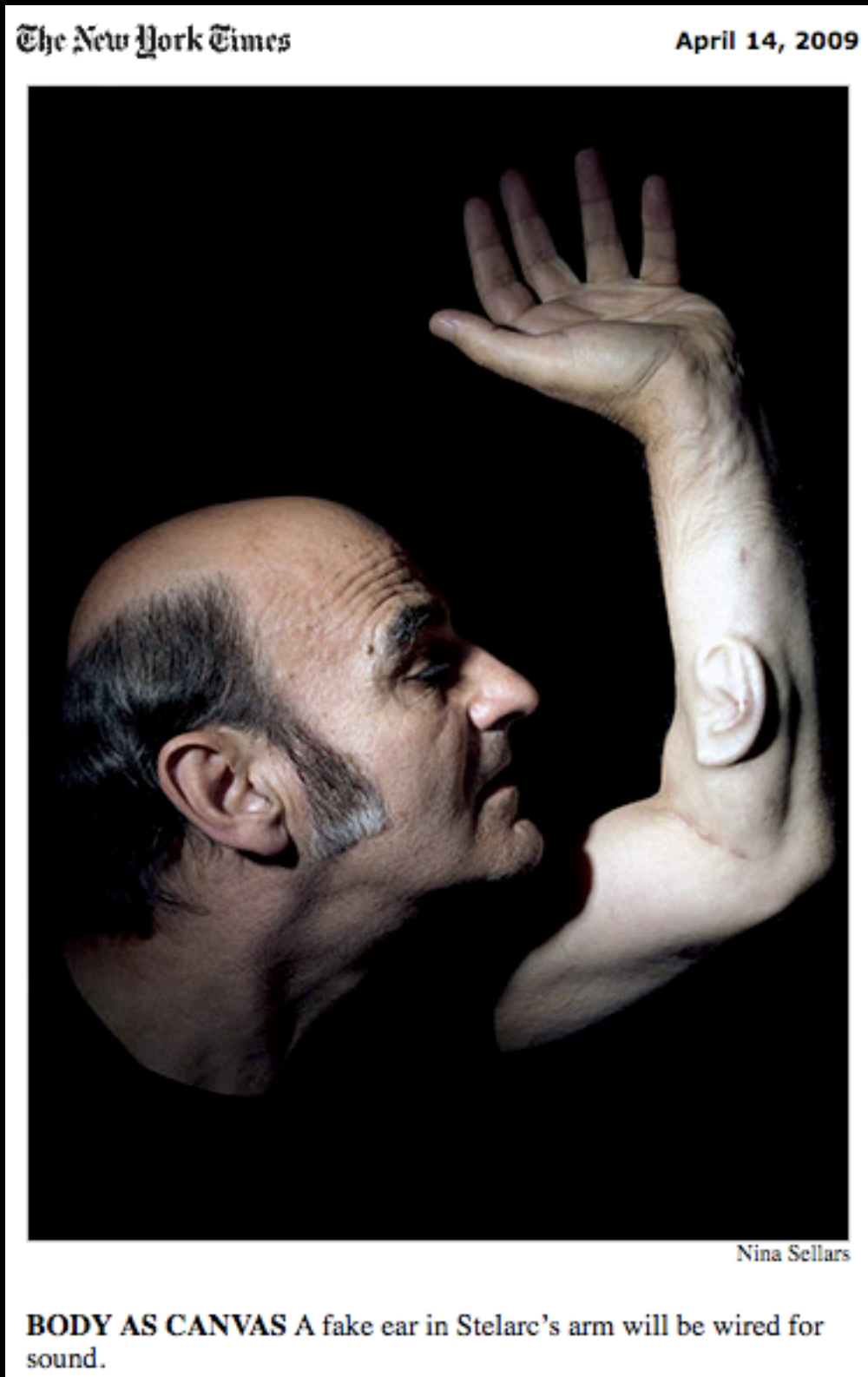
<http://www.nytimes.com/2009/04/14/science/14corp.html>

The **humanity** of synthetic biology



[http://archives.volitionwatch.com/download/fs I mod/4assmonkey.jpg](http://archives.volitionwatch.com/download/fs%20mod/4assmonkey.jpg)

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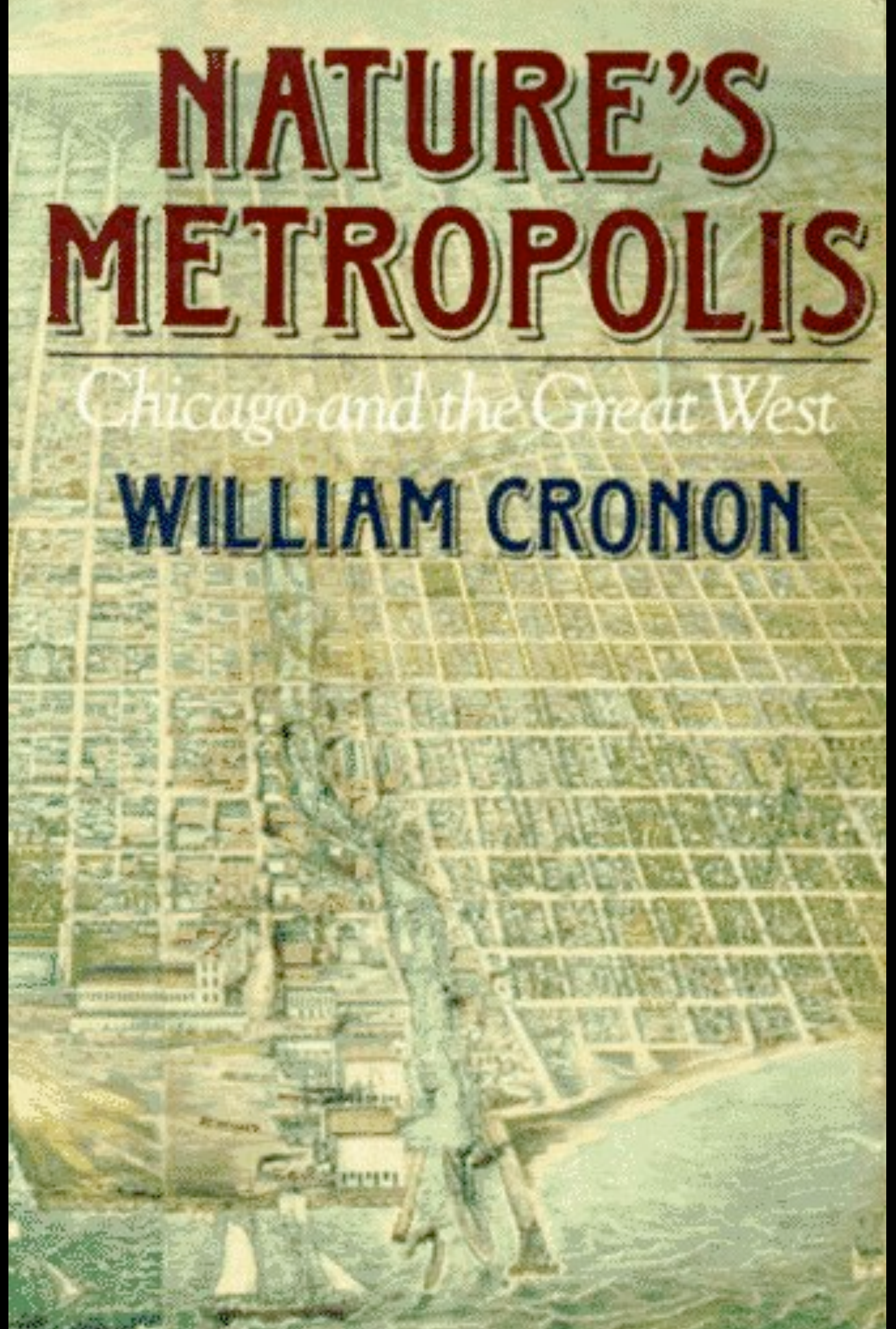


<http://s3.amazonaws.com/listverse/scifiseries/rama.jpg>

NATURE'S METROPOLIS

Chicago and the Great West

WILLIAM CRONON

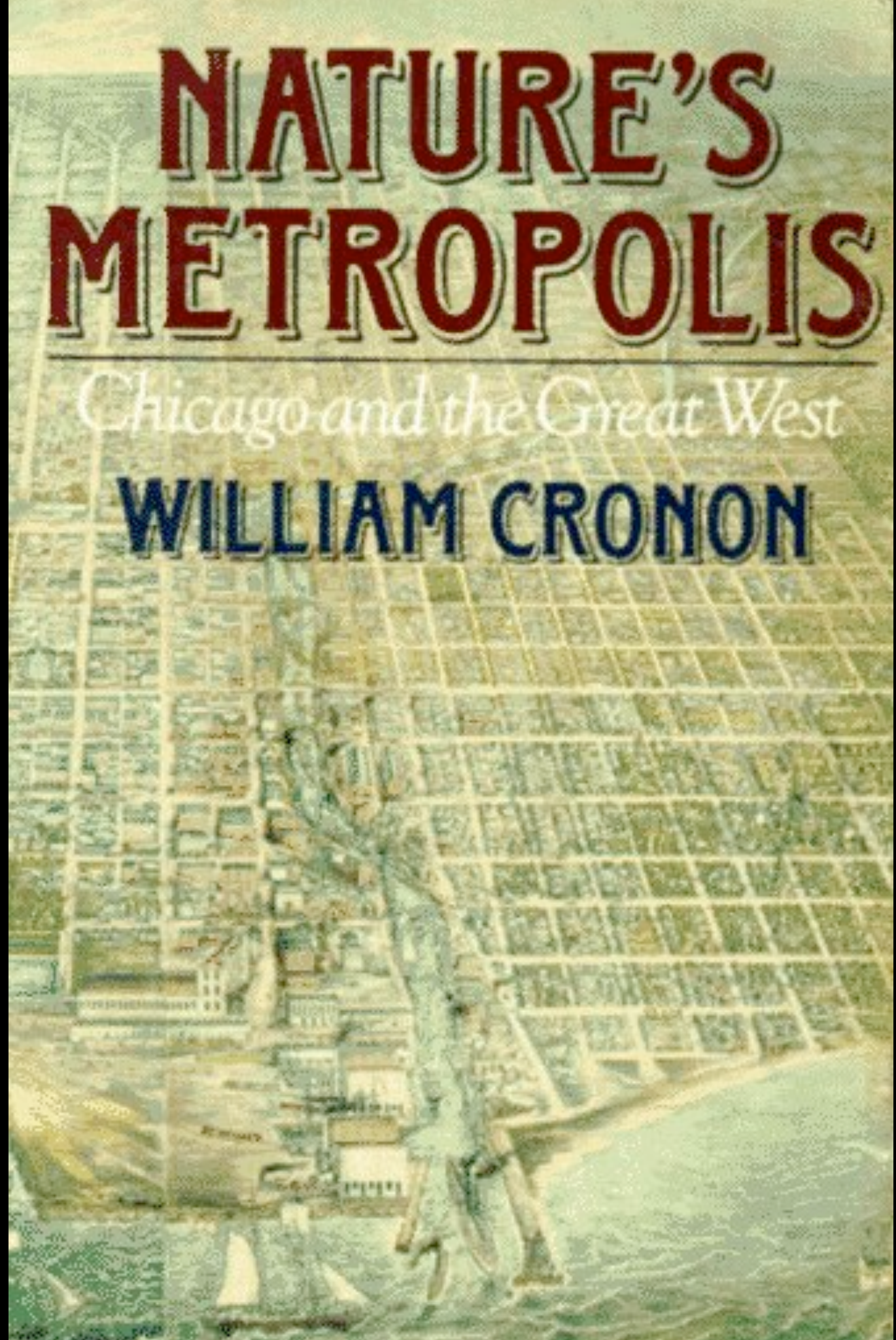


- How will (should?) we change ourselves and our environments?

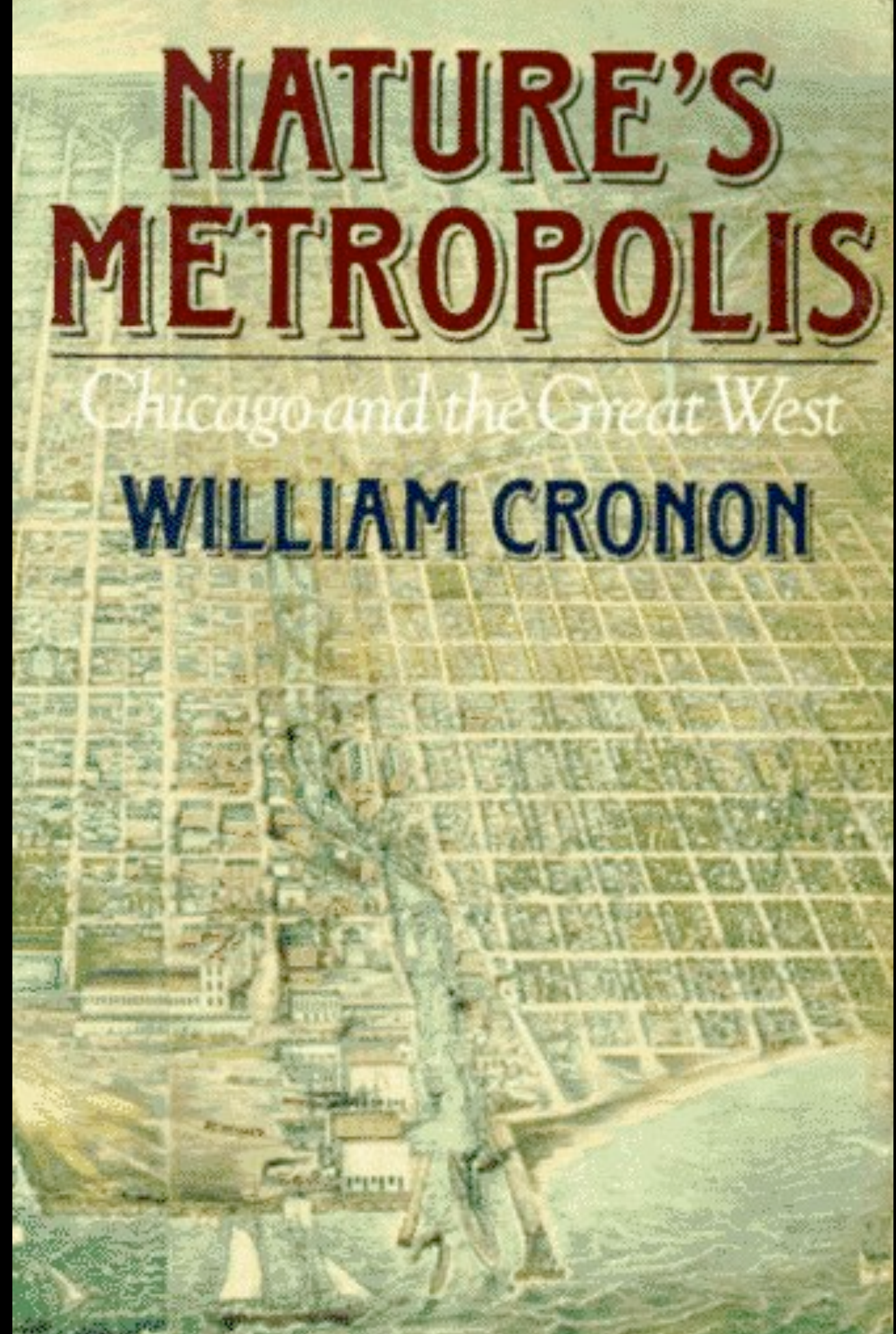
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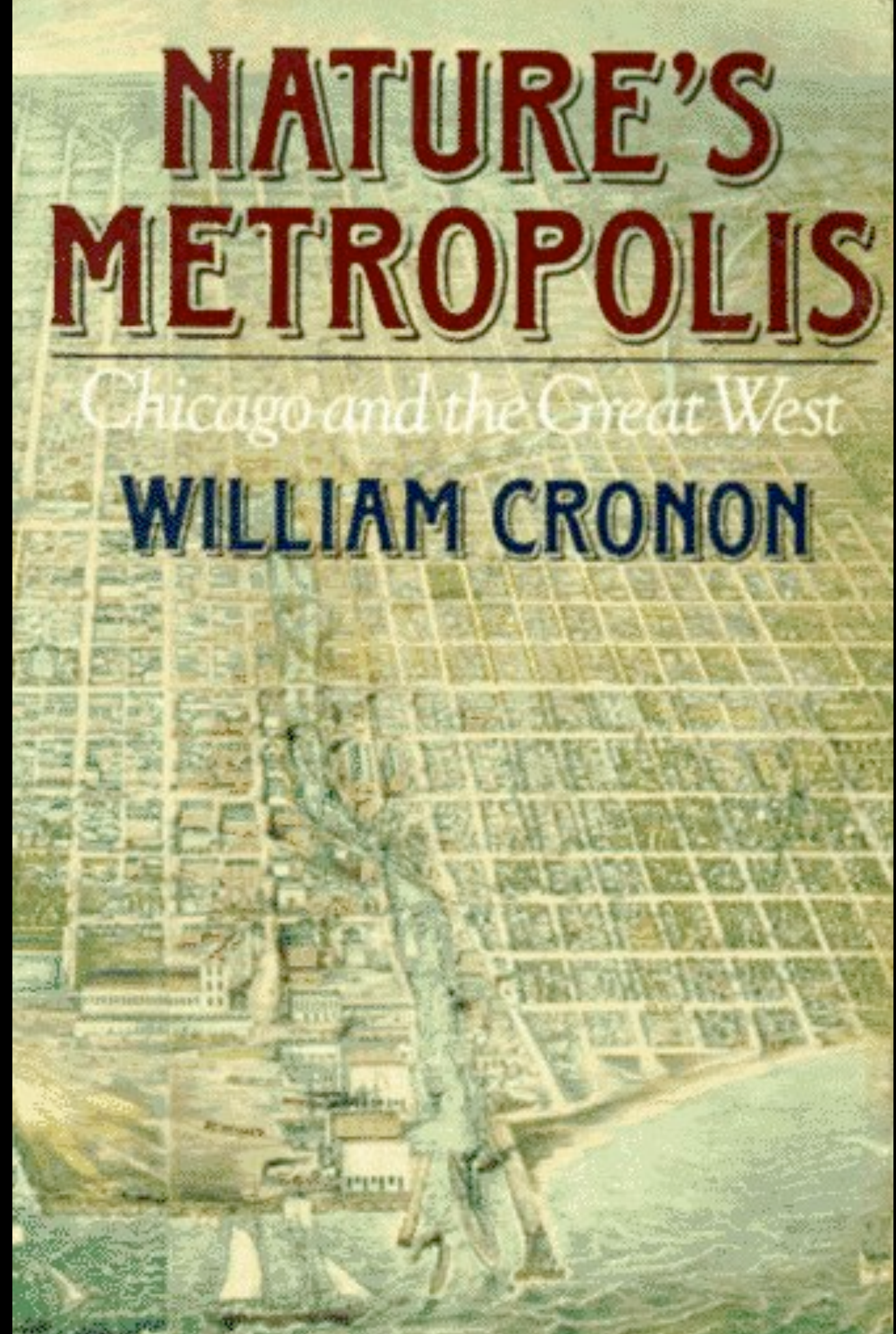
WILLIAM CRONON



- How will (should?) we change ourselves and our environments?
- New schools of science and engineering? E.g., integration without isolation?



- How will (should?) we change ourselves and our environments?
- New schools of science and engineering? E.g., integration without isolation?
- New modes of humanity? E.g., responsibility with representation?



Learn (& play) by making

Help (& enable) by building





Challenges

*Make biology easy
to engineer.*

Enable humanity.



Challenges

*Make biology easy
to engineer.*

Enable humanity.

Opportunities

*Enable all constructive
biotechnologies.*

*Better understand
nature.*

Food

Energy

Environment

Agriculture

Health

Chemicals

Security

Food

Energy

Environment

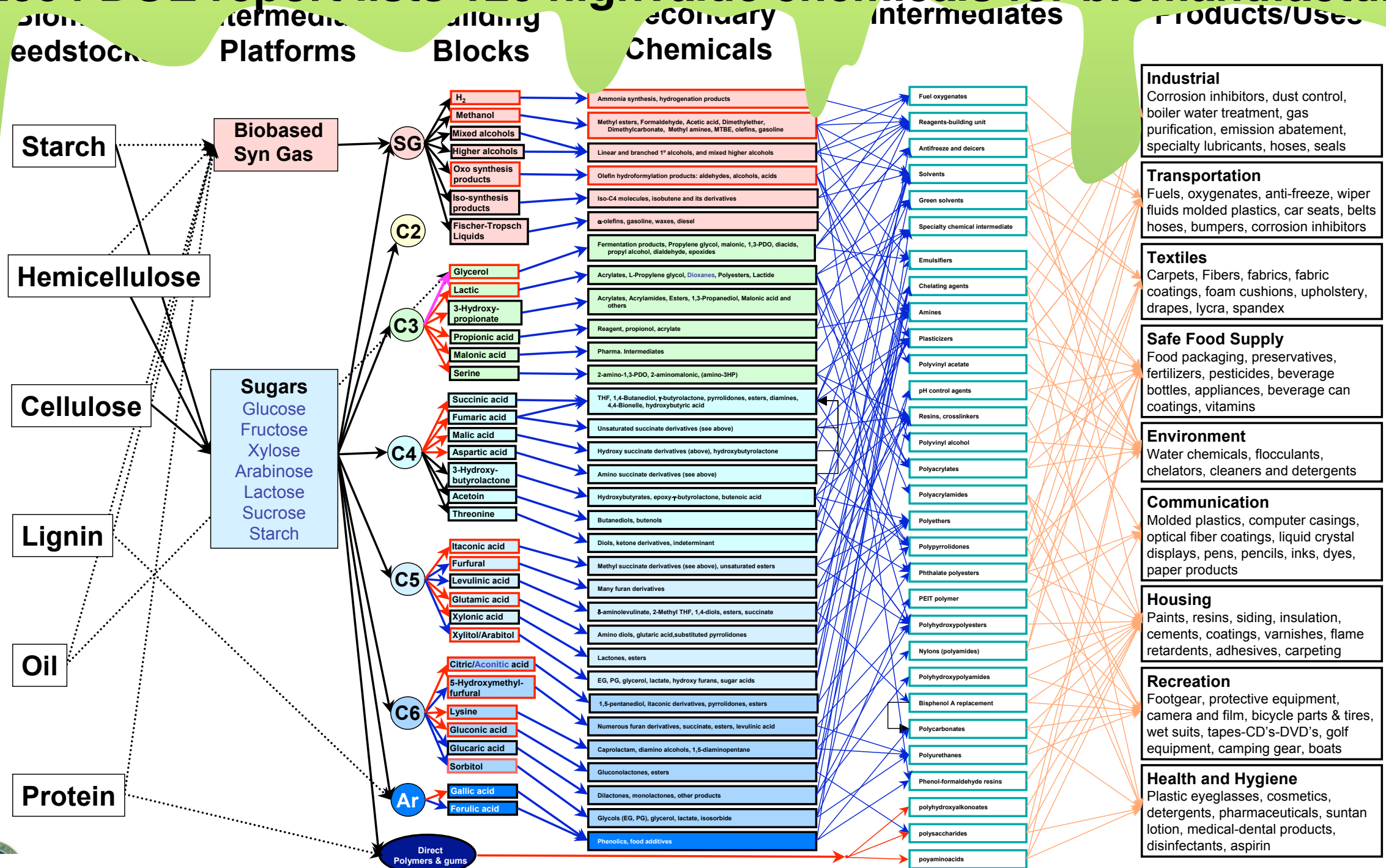
Agriculture

Health

Chemicals

Security

2004 DOE report lists 120 highvalue chemicals for biomanufacturing



Food

Energy

Environment

Agriculture

Health

Chemicals

Security

Food

Energy

Environment

Agriculture

Health

Chemicals

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Doebley Lab
Department of Genetics
University of Wisconsin-Madison

Food

Energy

Environment

Agriculture

Health

Chemicals

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3 crop species (rice, wheat and maize) provide 60% of all calories and 54% of all protein in human food

Doebly Lab
Department of Genetics
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Energy

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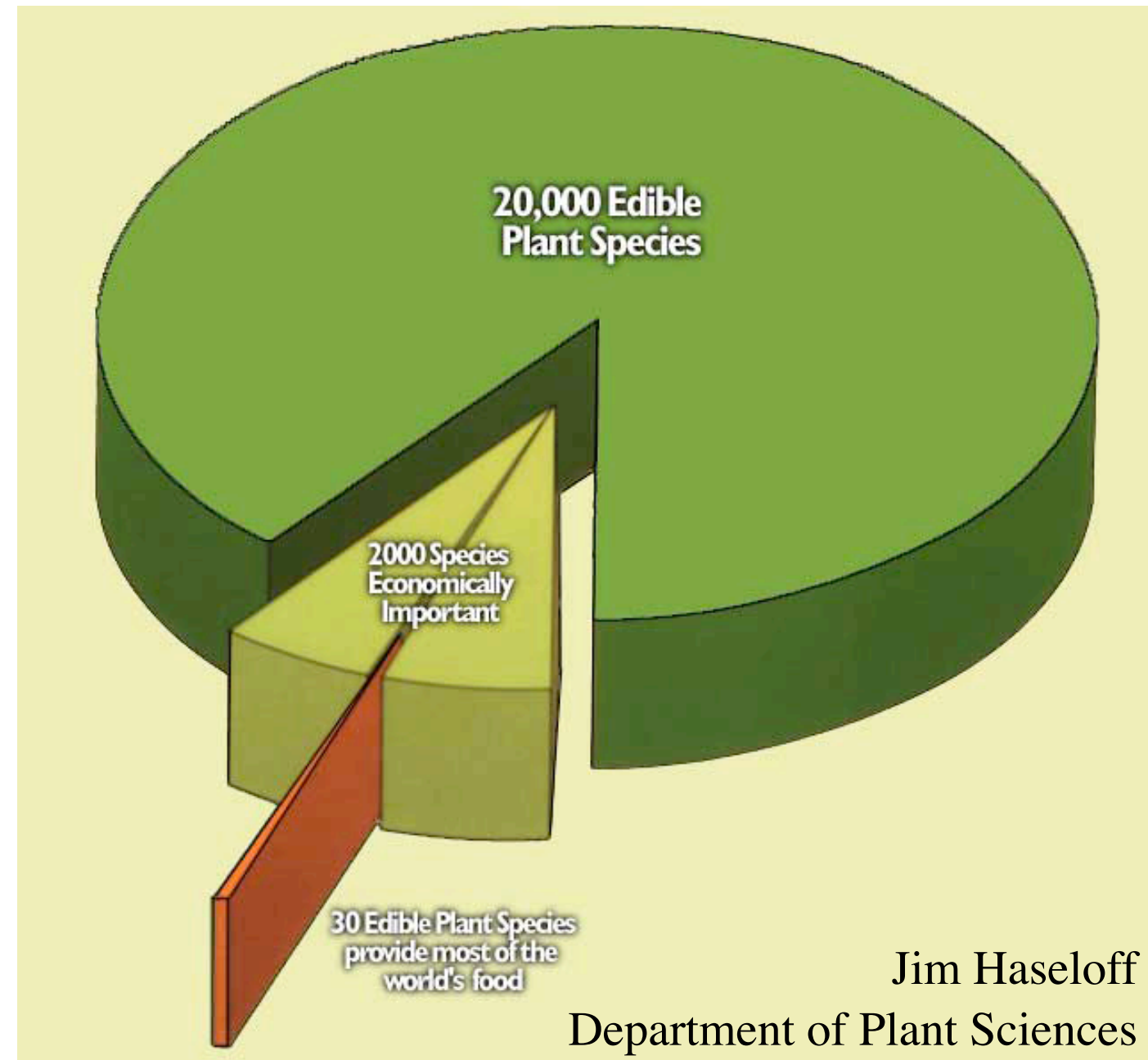
Health

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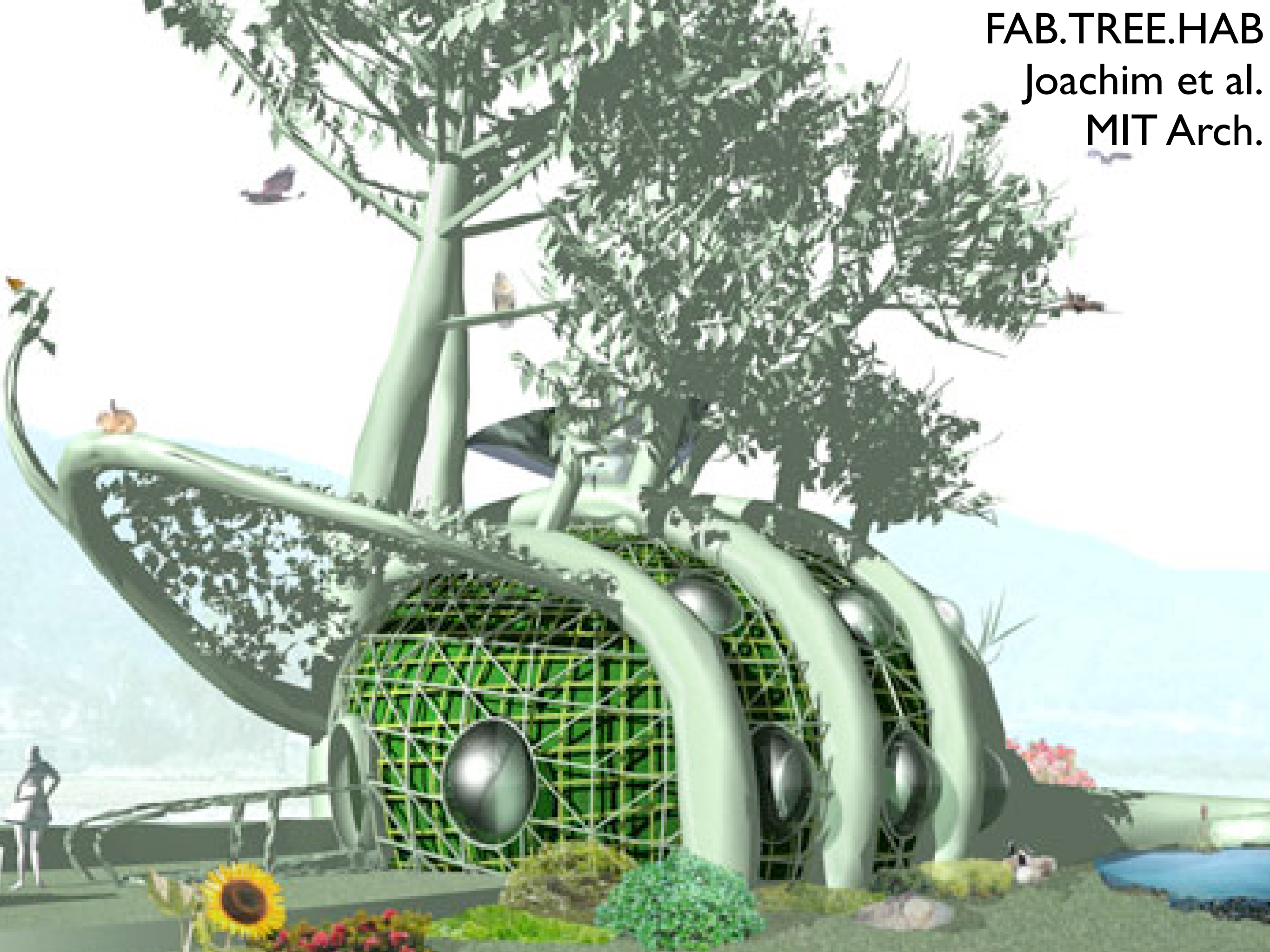
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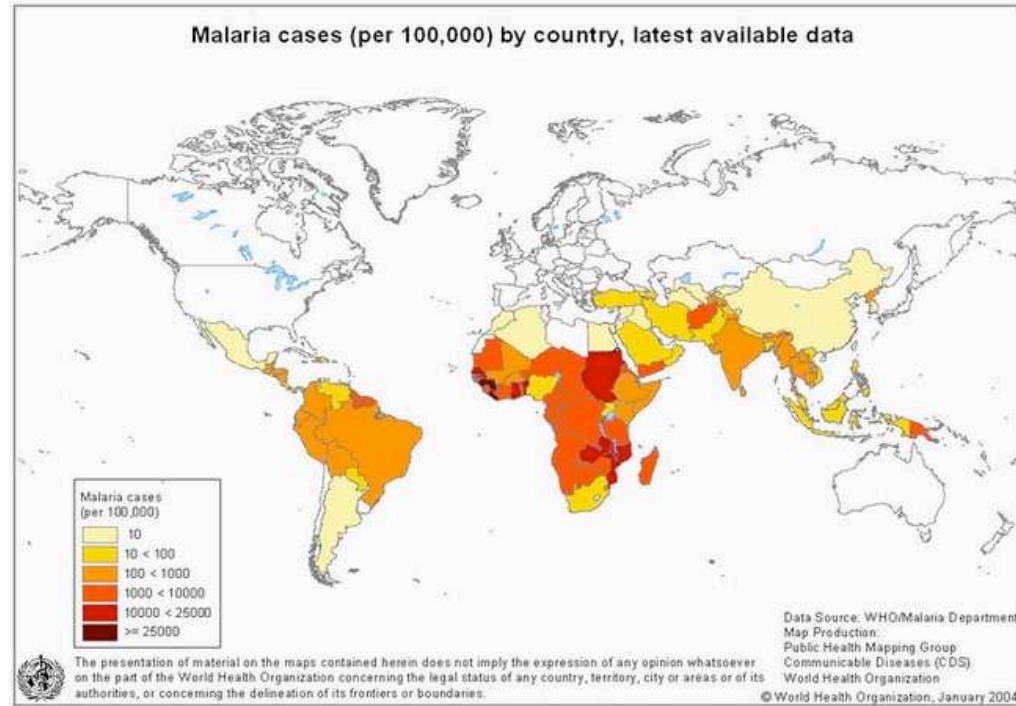
Doebley Lab
Department of Genetics
University of Wisconsin-Madison

Jim Haseloff
Department of Plant Sciences
University of Cambridge

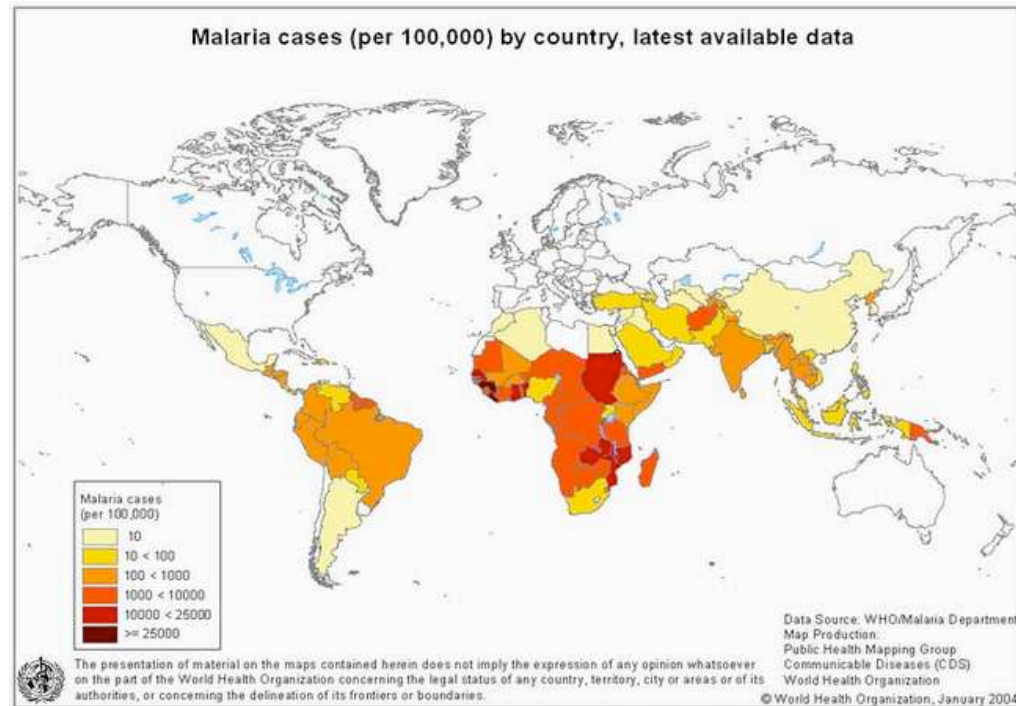
FAB.TREE.HAB
Joachim et al.
MIT Arch.



Where are we **today**?



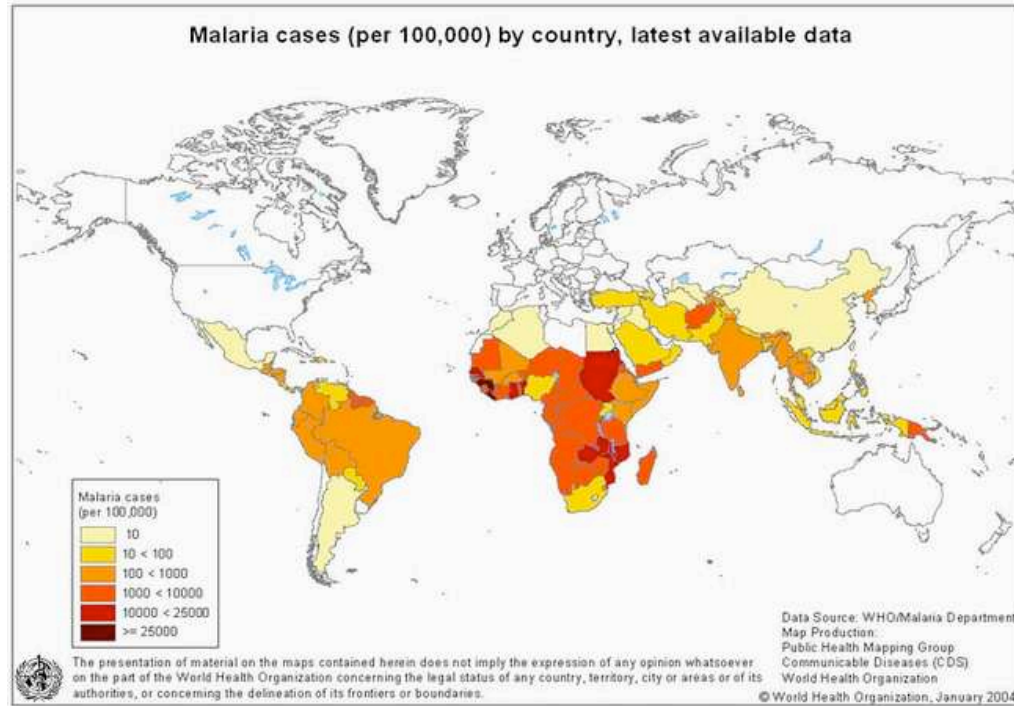
I) Malaria is a global problem, artemisinin offers a cure.



2) Jay Keasling's team spent \$25M to make artemisinin via biotechnology.

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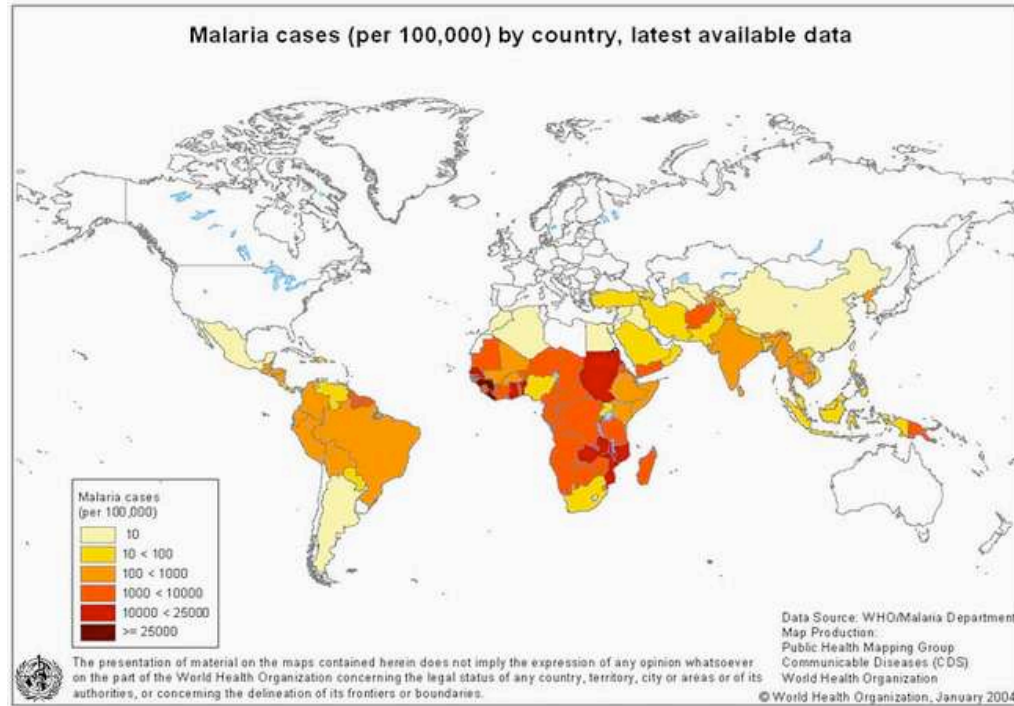
Today, each project is Herculean



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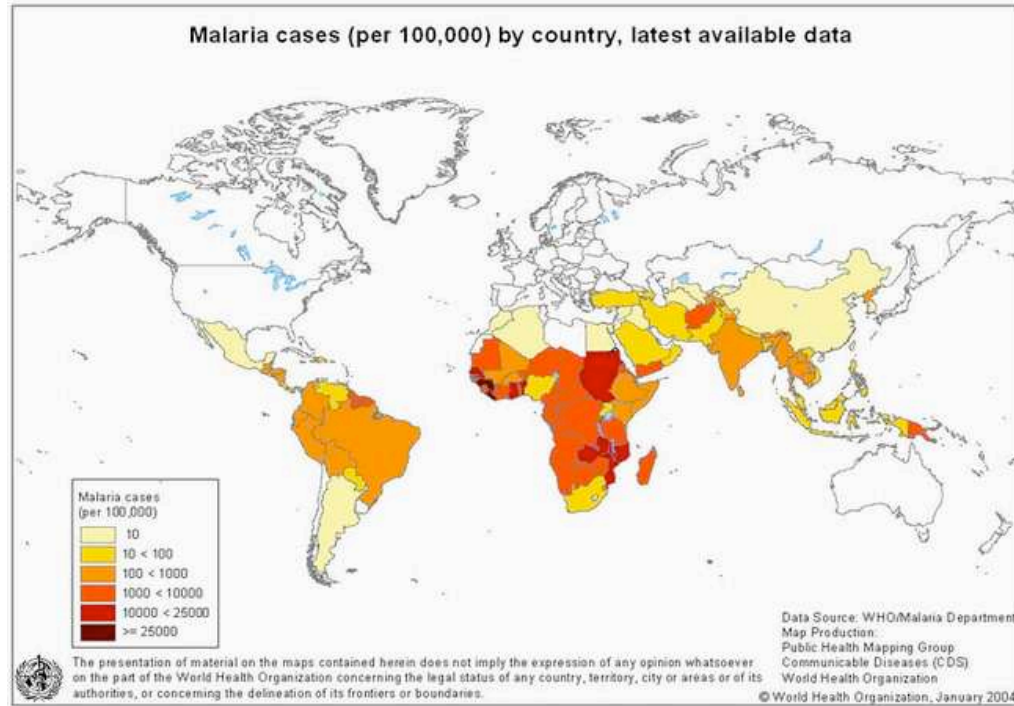
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3) But artemisinin resistance is already occurring.

Today, each project is Herculean



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3) But artemisinin resistance is already occurring.

Must we always spend many years and \$25M for each pressing biotech project?



Unknown
Our Lord's Candle (*Yucca whipplei*), Lakeview Mountains, 22 Apr 2006
Smugmug.com



Unknown

<http://www.rootsweb.com/~usgenweb/mn/stearns/postcards/quarry.jpg>

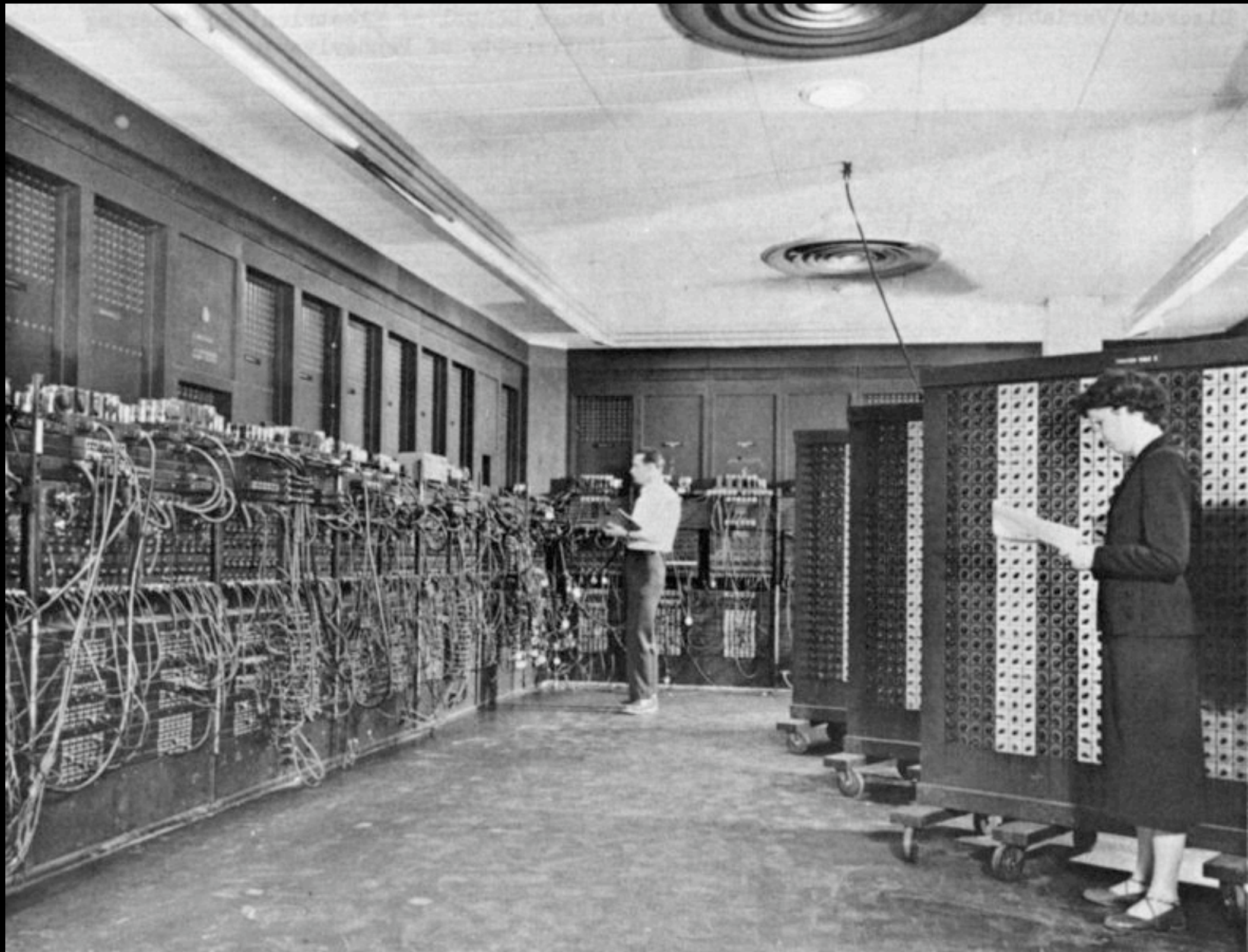


Washington National Cathedral

<http://www.cathedral.org/cathedral/discover/1930photos/5.shtml>

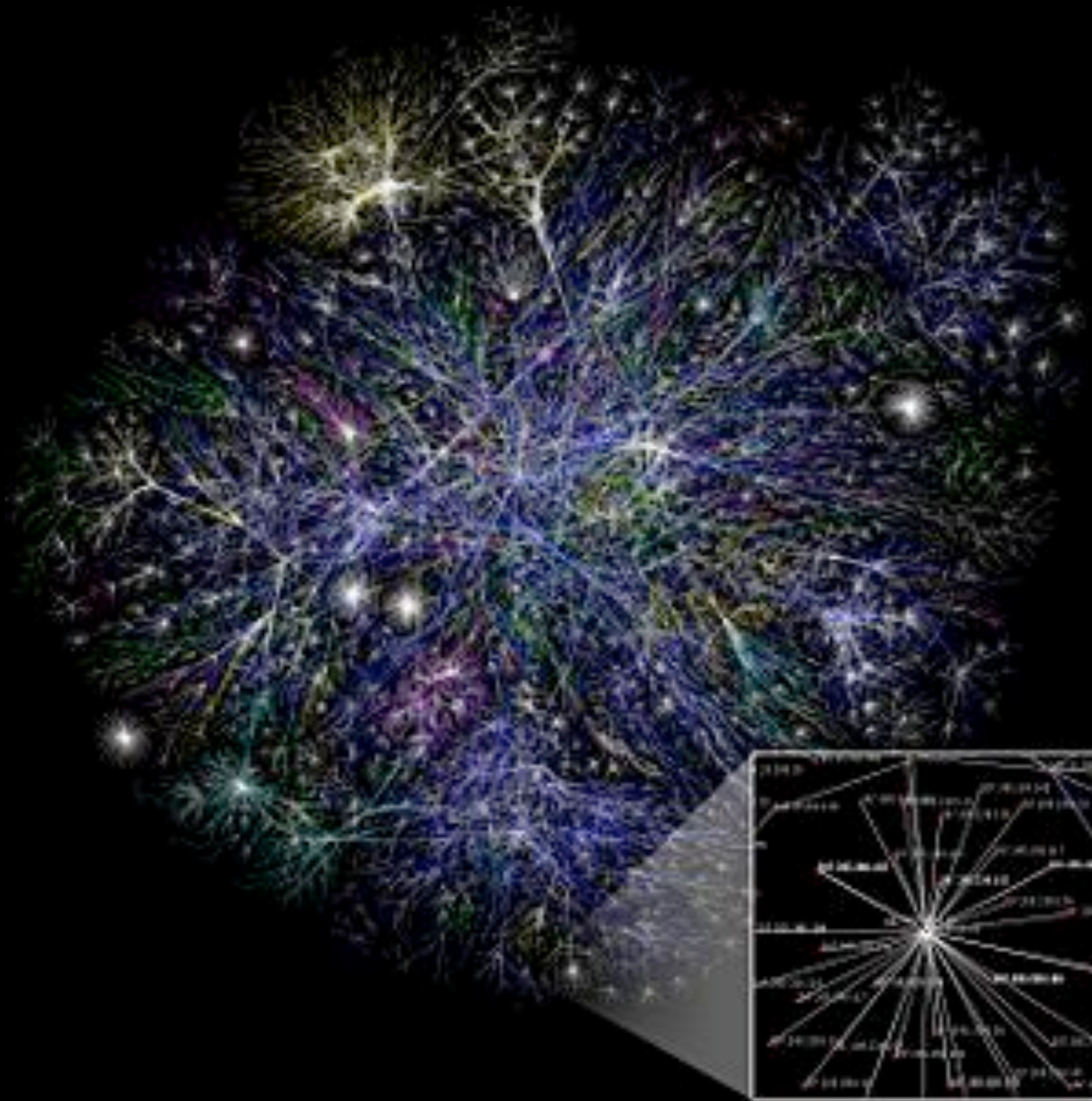








http://en.wikipedia.org/wiki/Image:Apple_I.jpg



<http://en.wikipedia.org/wiki/Internet>

A *p p* S

Tools

I. Info. theory & signal proc.

7. Control & dyn. systems
6. Reverse engineering
5. Fab, CAD & EDA
4. Standards & abstraction
3. Languages & grammars
2. Device design
1. Info. theory & signal proc.

1973

Construction of biologically functional bacterial plasmids in vitro

Cohen et al., PNAS, 1973

MATERIALS AND METHODS

E. coli strain W1485 containing the RSF1010 plasmid, which carries resistance to streptomycin and sulfonamide, was obtained from S. Falkow. Other bacterial strains and R factors and procedures for DNA isolation, electron microscopy, and transformation of *E. coli* by plasmid DNA have been described (1, 7, 8). Purification and use of the *Eco*RI restriction endonuclease have been described (5). Plasmid heteroduplex studies were performed as previously described (9, 10). *E. coli* DNA ligase was a gift from P. Modrich and R. L. Lehman and was used as described (11). The detailed procedures for gel electrophoresis of DNA will be described elsewhere (Helling, Goodman, and Boyer, in preparation); in brief, duplex DNA was subjected to electrophoresis in a tube-type apparatus (Hoefer Scientific Instrument) (0.6 × 15-cm gel) at about 20° in 0.7% agarose at 22.5 V with 40 mM Tris-acetate buffer (pH 8.05) containing 20 mM sodium acetate, 2 mM EDTA, and 18 mM sodium chloride. The gels were then soaked in ethidium bromide (5 µg/ml) and the DNA was visualized by fluorescence under long wavelength ultraviolet light ("black light"). The molecular weight of each fragment in the range of 1 to 200 × 10⁶ was determined from its mobility relative to the mobilities of DNA standards of known molecular weight included in the same gel (Helling, Goodman, and Boyer, in preparation).

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1985

Cloning and expression of the human erythropoietin gene

Lin et al., PNAS, 1985

Assembly of Expression Vector for the Epo Gene. For direct expression of the genomic Epo gene, the 4.8-kilobase (kb) *Bst*EII-*Bam*HI fragment of λHE1 (see *Results*), which contains the entire Epo gene, was used. After converting the *Bst*EII site into a *Bam*HI site with a synthetic linker, the fragment was inserted into the unique *Bam*HI site of the expression vector pDSVL (unpublished data), which contains a dihydrofolate reductase (DHFR) minigene from pMg1 (24). The resulting plasmid pDSVL-gHuEPO (Fig. 1A) was then used to transfect Chinese hamster ovary (CHO) DHFR⁻ cells (25) by the calcium phosphate microprecipitate method (26). The transformants were selected by growth in medium lacking hypoxanthine and thymidine. The culture medium used was Dulbecco's modified Eagle's medium supplemented with 10% fetal bovine serum, penicillin, streptomycin, and glutamine (25).

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Assembly of Expression Vector for the Epo Gene. For direct expression of the genomic Epo gene, the 4.8-kilobase (kb) *Bst*EII-*Bam*HI fragment of λHE1 (see *Results*), which contains the entire Epo gene, was used. After converting the *Bst*EII site into a *Bam*HI site with a synthetic linker, the fragment was inserted into the unique *Bam*HI site of the expression vector pDSVL (unpublished data), which contains a dihydrofolate reductase (DHFR) minigene from pMg1 (24). The resulting plasmid pDSVL-gHuEPO (Fig. 1A) was then used to transfect Chinese hamster ovary (CHO) DHFR⁻ cells (25) by the calcium phosphate microprecipitate method (26). The transformants were selected by growth in medium lacking hypoxanthine and thymidine. The culture medium used was Dulbecco's modified Eagle's medium supplemented with 10% fetal bovine serum, penicillin, streptomycin, and glutamine (25).

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For integration of an expression cassette for *tHMGR*, plasmid pδ-HMGR was constructed. First *Sac*II restriction sites were introduced into pRS426GAL1¹⁰ at the 5' end of the *GAL*I promoter and 3' end of the *CYC*I terminator. To achieve this, the promoter-multiple cloning site-terminator cassette of pRS426GAL1 was PCR amplified using primer pair 11 and 12. The amplified product was cloned directly into *Pvu*II-digested pRS426GAL1 to construct vector pRS426-SacII. The catalytic domain of *HMGR* was PCR amplified from plasmid pRH127-3¹¹ with primer pair 13 and 14. The amplified product was cleaved with *Bam*HI and *Sac*II and cloned into *Bam*HI and *Xho*I digested pRS426-SacII. pRS-HMGR was cleaved with *Sac*II and the expression

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Construction of biologically functional bacterial plasmids in vitro

Cohen et al., PNAS, 1973

MATERIALS AND METHODS
E. coli strain W1485 containing the RSF1010 plasmid, which carries resistance to streptomycin and sulfonamide, was obtained from S. Falkow. Other bacterial strains and R factors and procedures for DNA isolation, electron microscopy, and transformation of *E. coli* by plasmid DNA have been described (1, 7, 8). Purification and use of the *EcoRI* restriction endonuclease have been described (5). Plasmid heteroduplex studies were performed as previously described (9, 10). *E. coli* DNA ligase was a gift from P. Modrich and R. L. Lehman and was used as described (11). The detailed procedures for gel electrophoresis of DNA will be described elsewhere (Helling, Goodman, and Boyer, in preparation); in brief, duplex DNA was subjected to electrophoresis in a tube-type apparatus (Hoefer Scientific Instrument) (0.6 × 15-cm gel) at about 20° in 0.7% agarose at 22.5 V with 40 mM Tris-acetate buffer (pH 8.05) containing 20 mM sodium acetate, 2 mM EDTA, and 18 mM sodium chloride. The gels were then soaked in ethidium bromide (5 µg/ml) and the DNA was visualized by fluorescence under long wavelength ultraviolet light ("black light"). The molecular weight of each fragment in the range of 1 to 200 × 10³ was determined from its

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Genetic engineering basics unchanged past 30+ years

We need new tools

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Genetic engineering basics unchanged past 30+ years

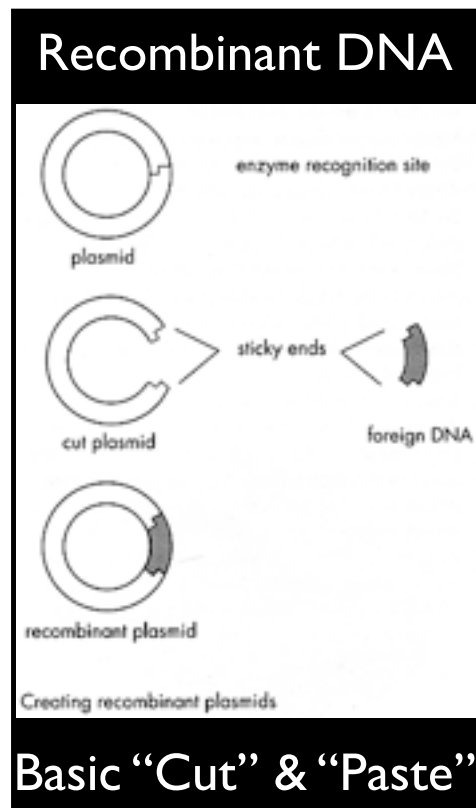
Synthetic Biology as Tools Revolution

Synthetic Biology as Tools Revolution

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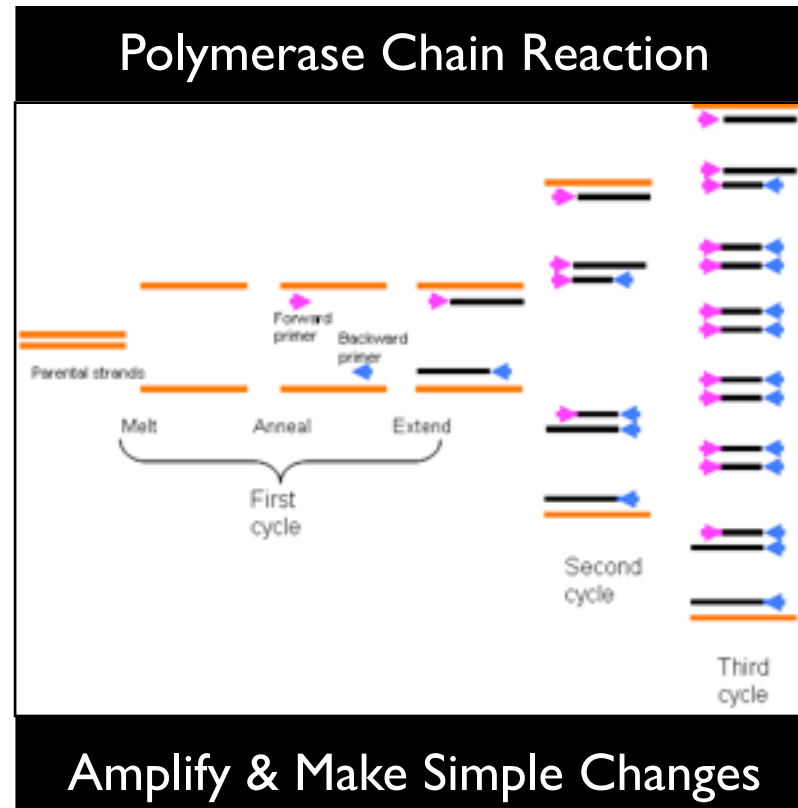
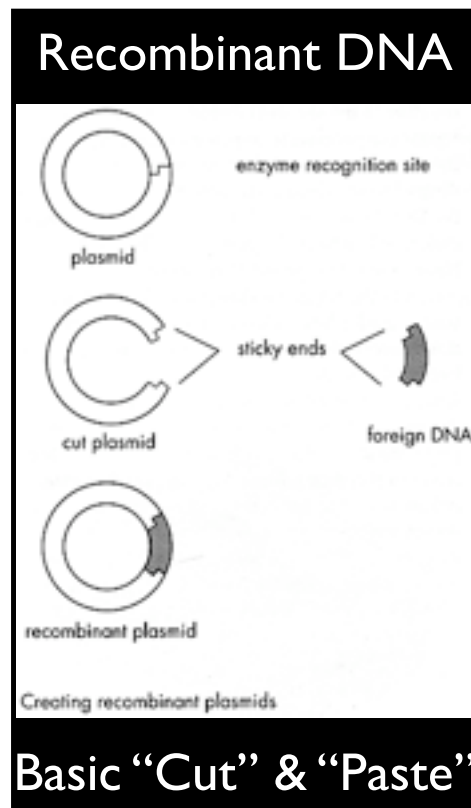
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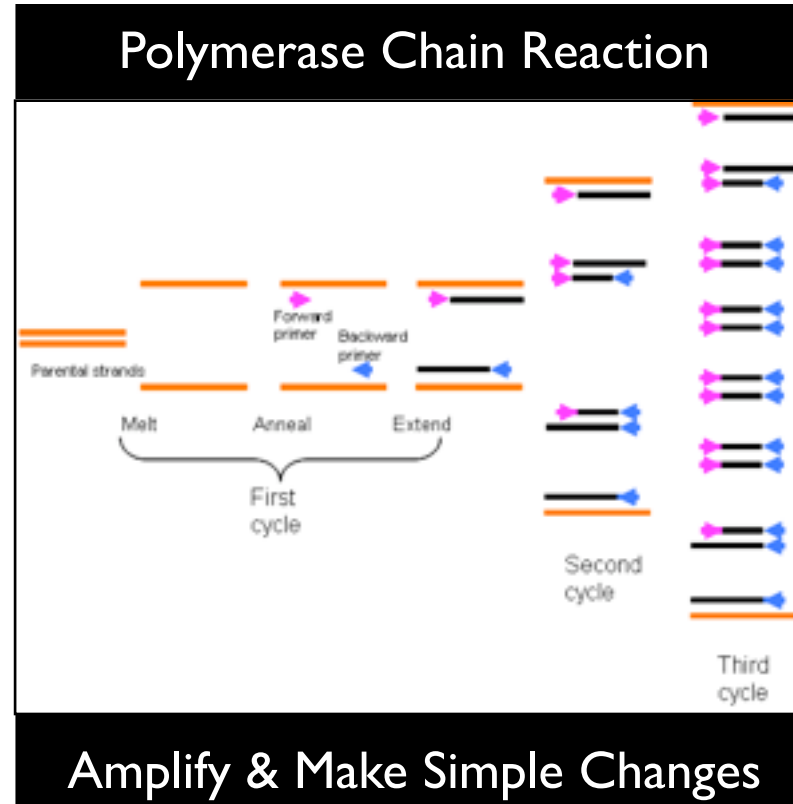
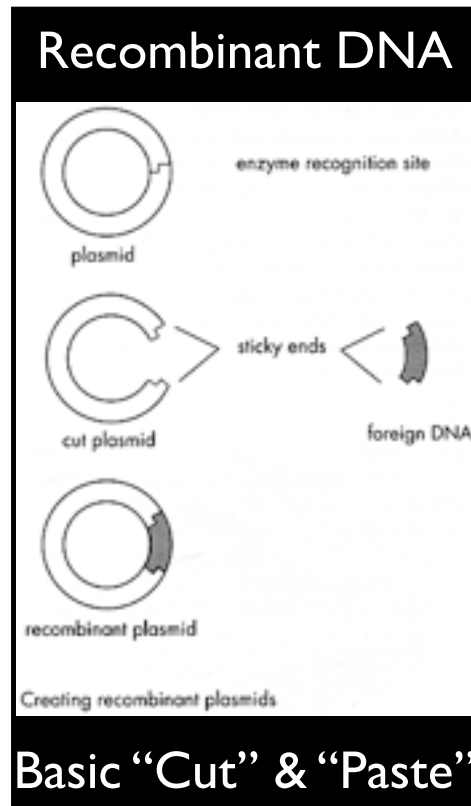
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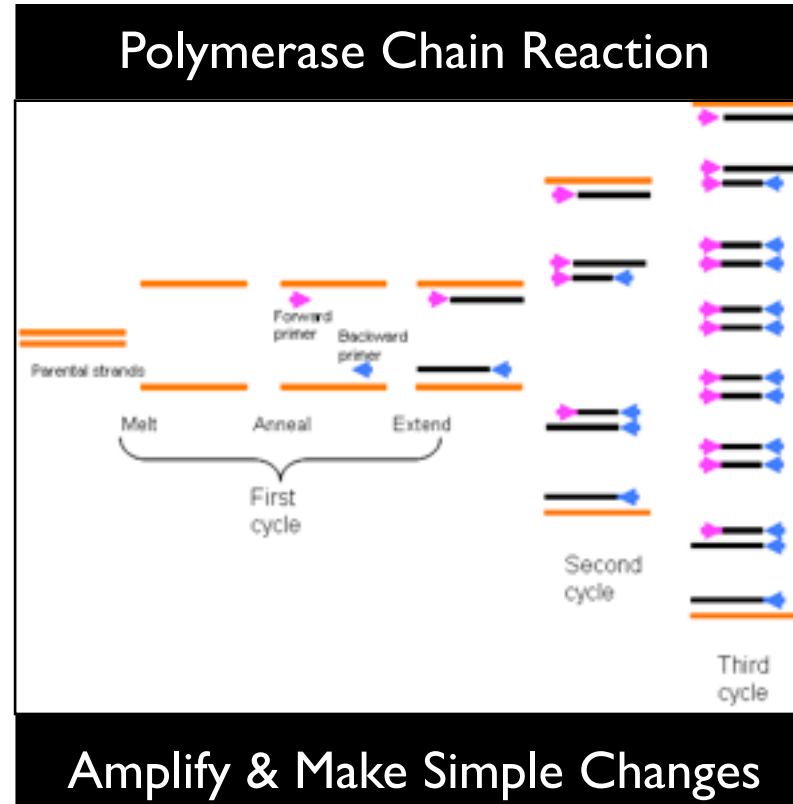
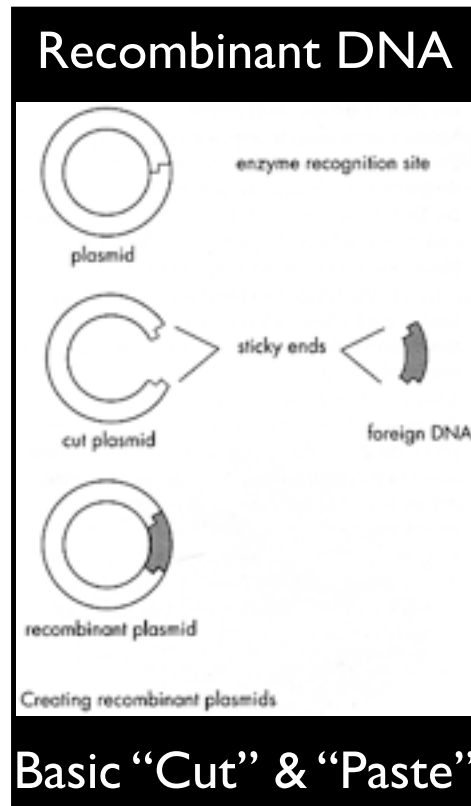
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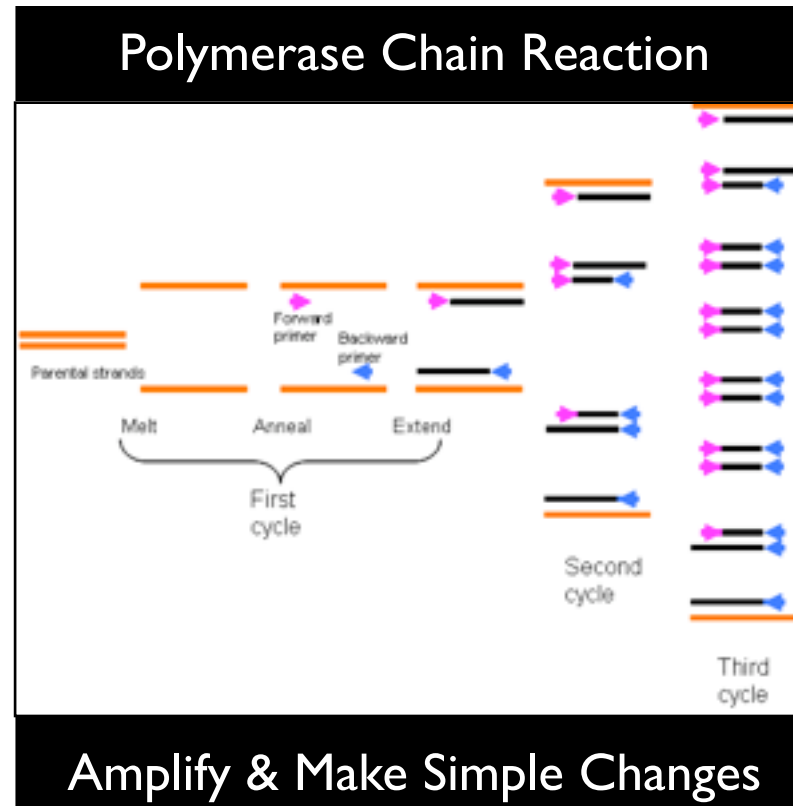
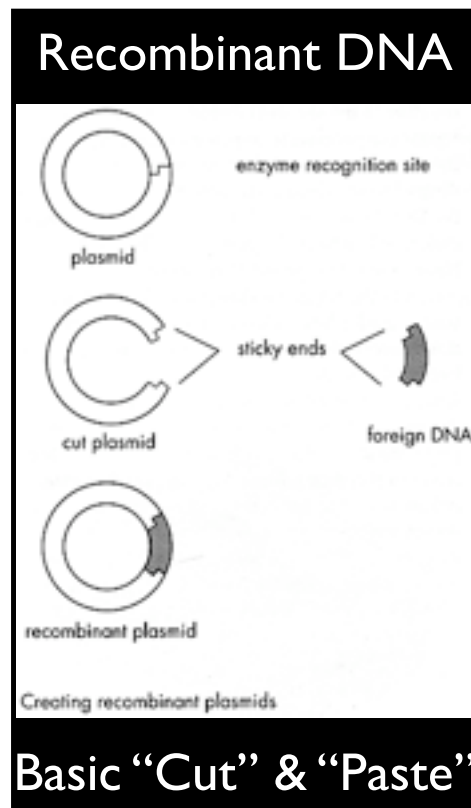
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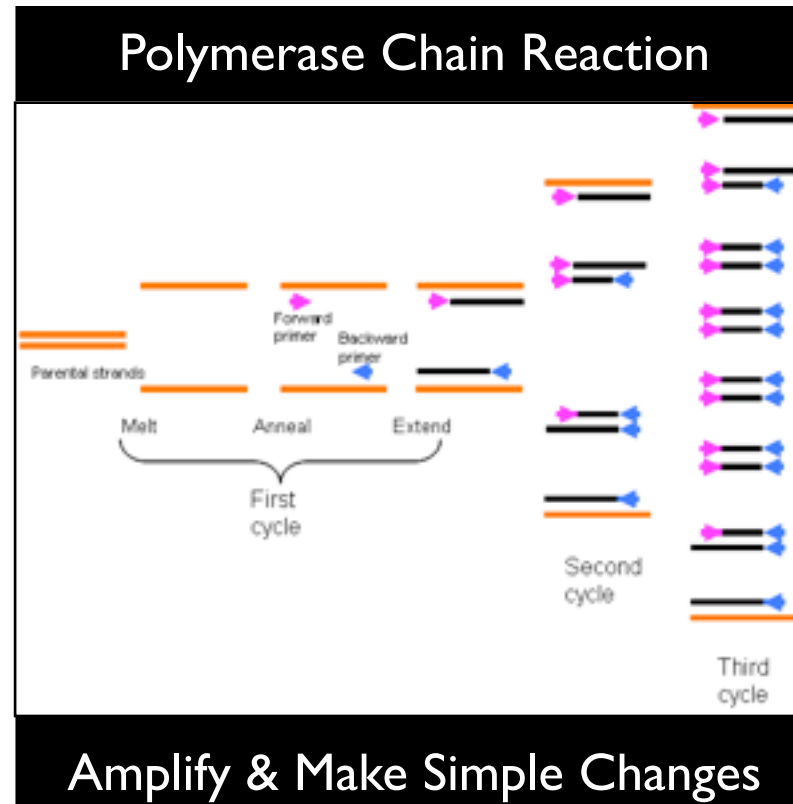
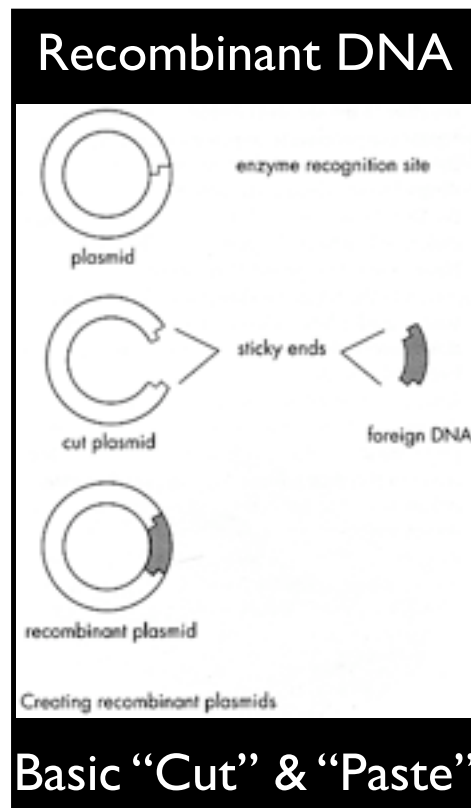


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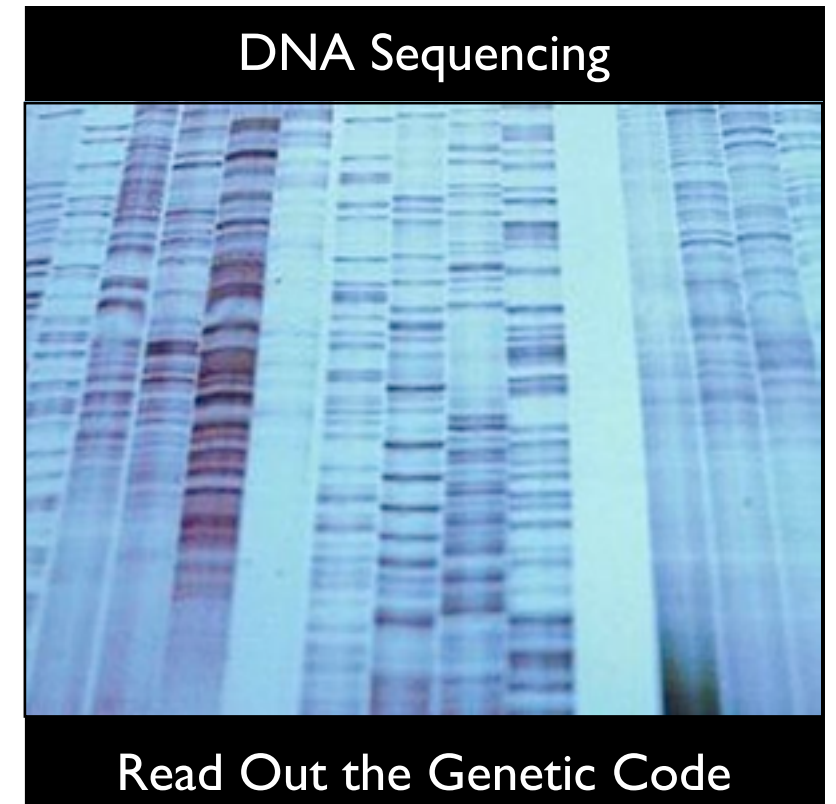
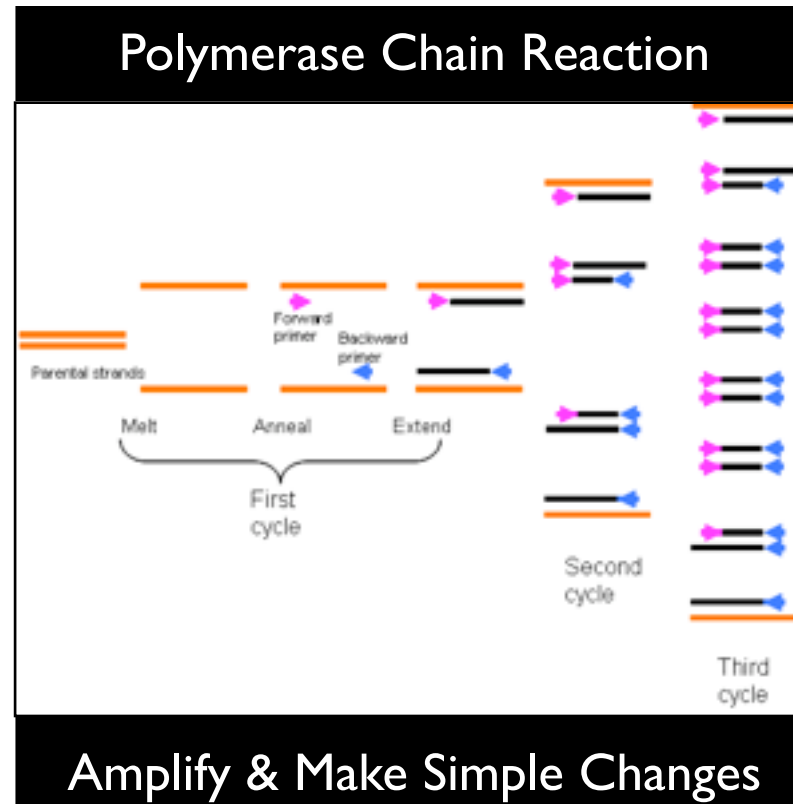
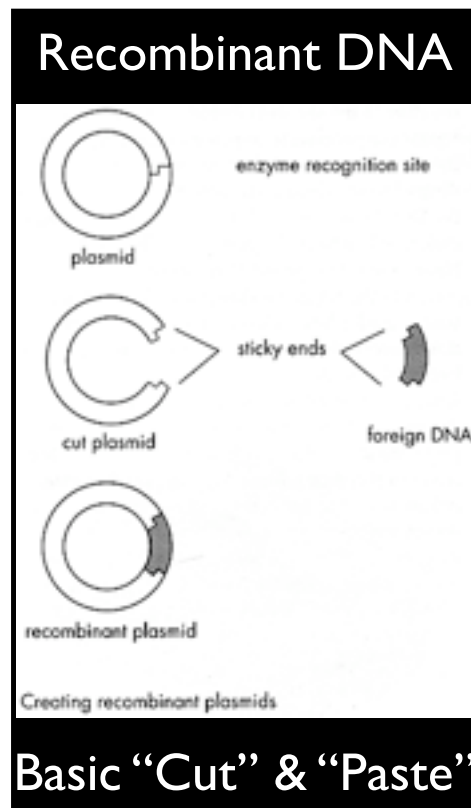


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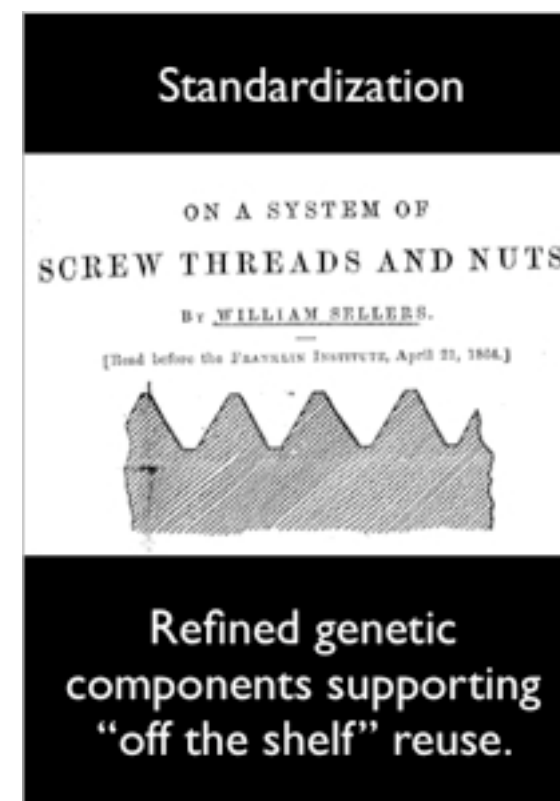


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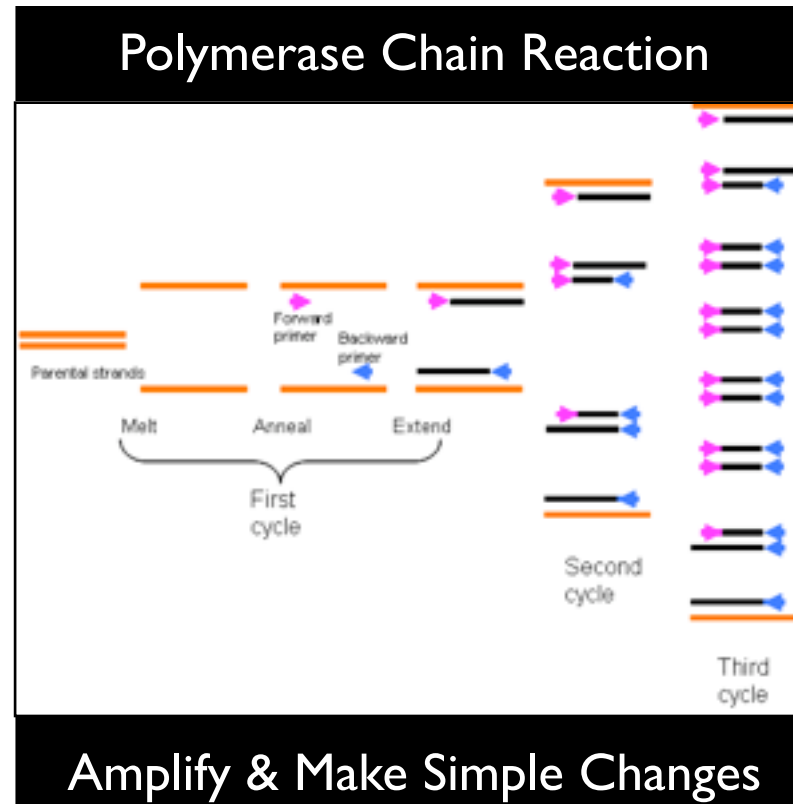
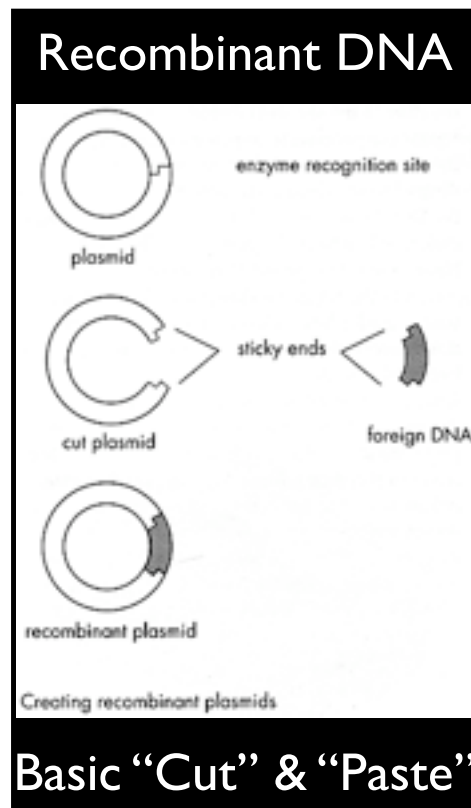


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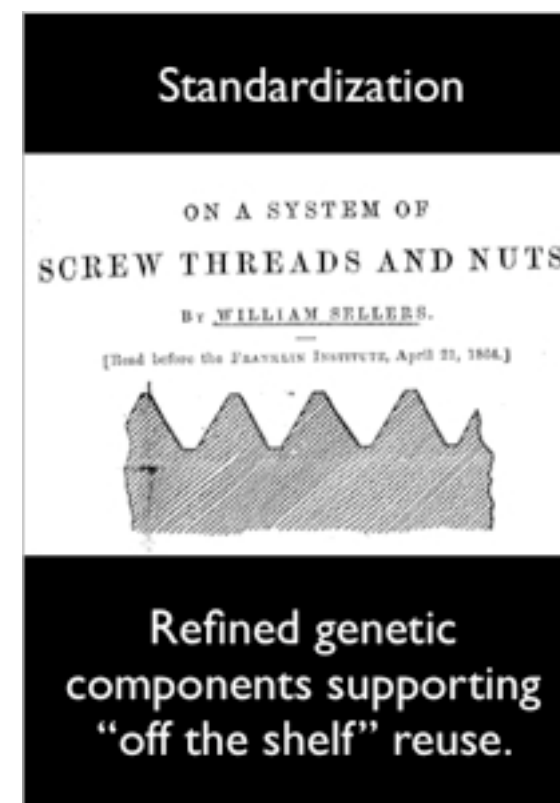


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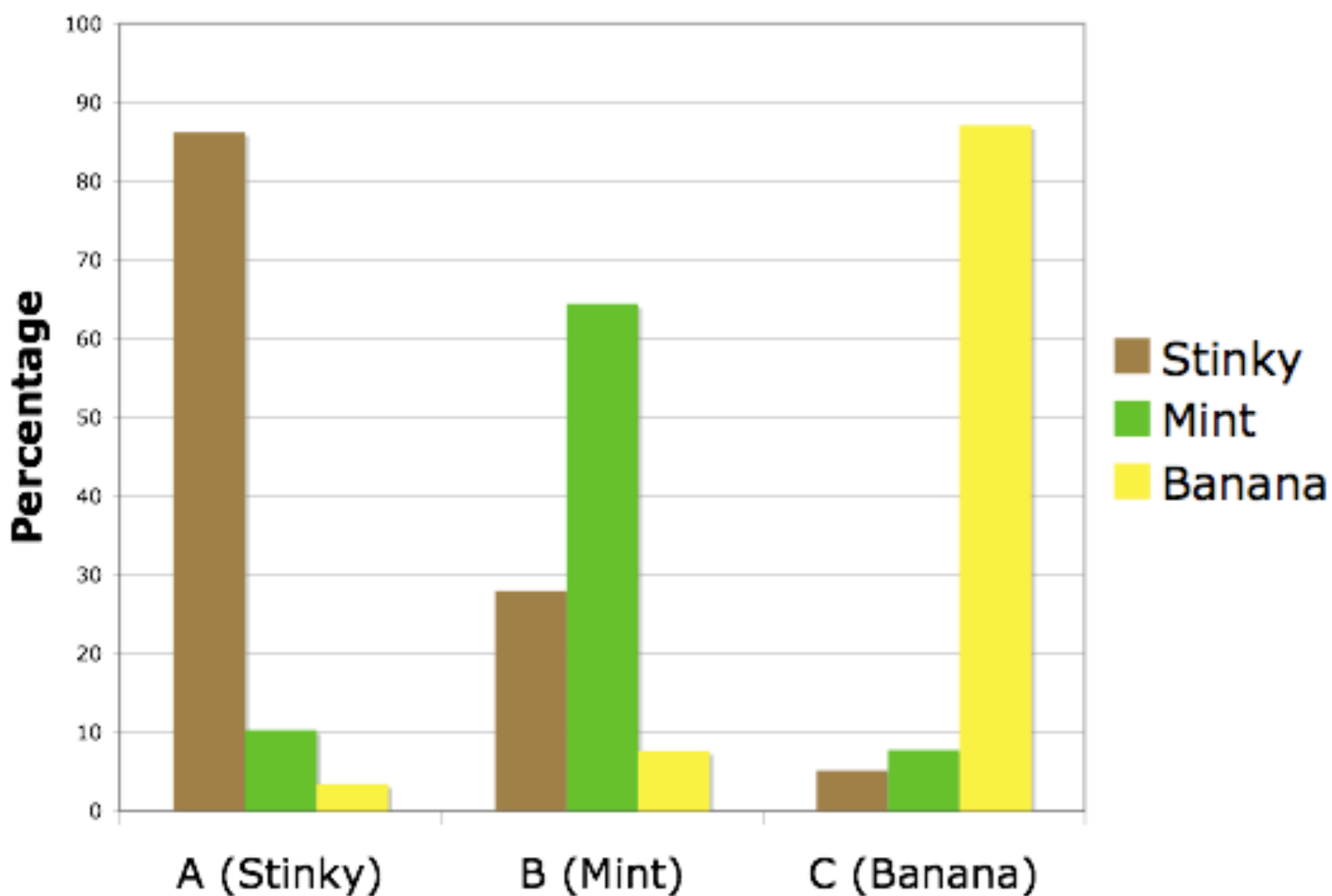


```
if {growing}  
    call wintergreen()  
else  
    call bananas()
```

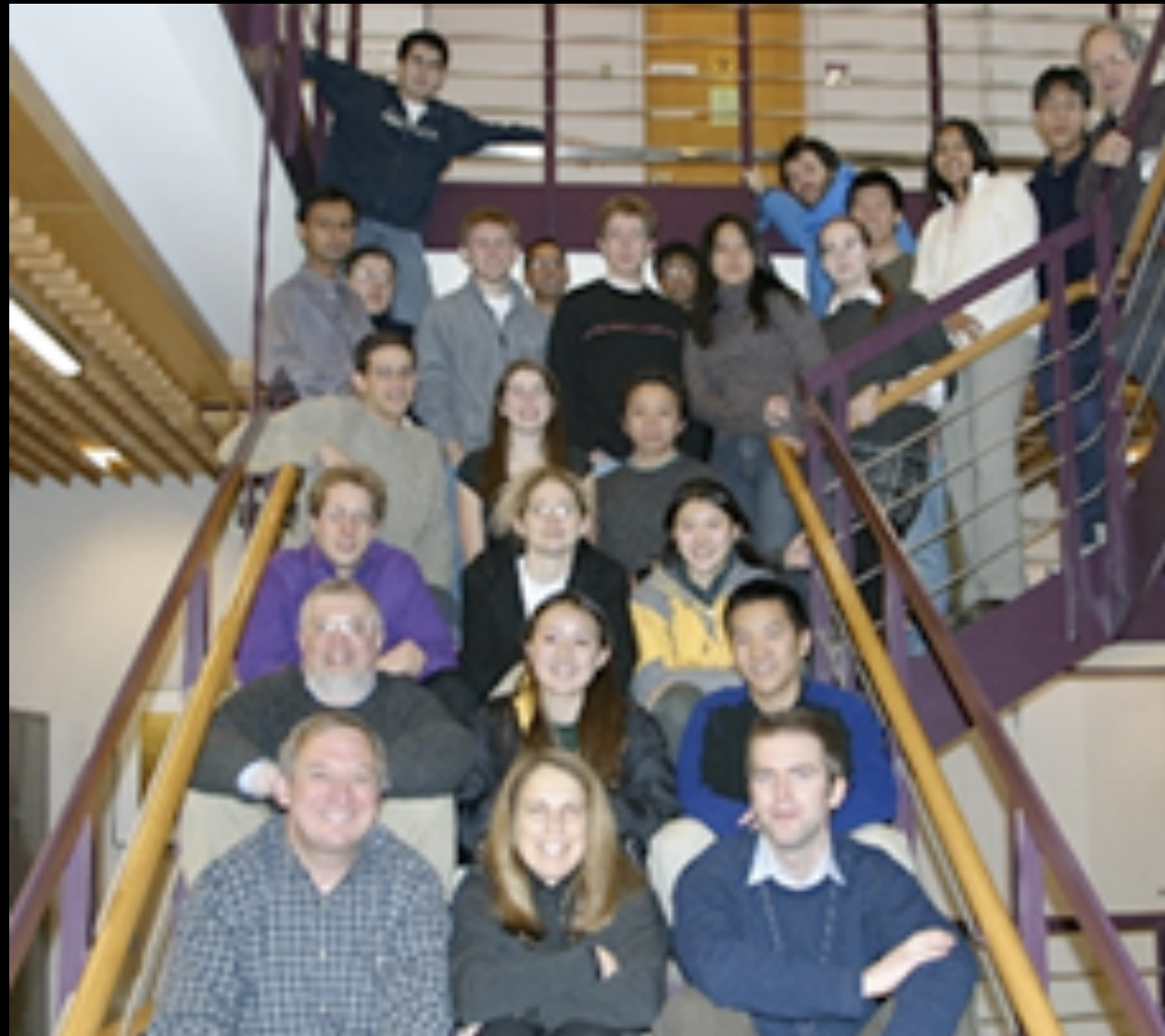

smell test: iGEM jamboree



smell test results















International Genetically Engineered Machine Competition

© J. R. Brown, iGEM 2006

Global Distribution of Competing Teams









Should teenagers practice genetic engineering?

Yes

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Yes

Should military force include biotechnology?

No

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Will biohackers be good or bad?

Good

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How much can we produce with biology?

???

The Imperative of Synthetic Biology: A Proposed National Research Initiative

Drew Endy
Stanford University

Ed Lazowska
University of Washington

Version 6: December 12, 2008¹

The oxygen in the air we breathe, the plants or animals we eat, the environments we cherish and struggle to protect, the fuels that keep us warm and power our industries and vehicles, our medicines and clothes, ourselves and future generations as yet unborn – we are of biology and dependant upon her. In turn, our ability to develop and deploy biology as a technology – for sustainable energy production, green manufacturing, agile crop development, affordable healthcare and medicines – depends on the tools we have for engineering life itself.

35 years ago researchers learned to directly manipulate DNA using crude molecular tools to construct relatively simple genetic programs. These first tools gave birth to the biotechnology industry, resulting in new drugs and therapies (e.g., from recombinant insulin for treating diabetes to cheap artemisinin for fighting malaria), concerns (e.g., biological security), controversies (e.g., genetically engineered foods), and unmet promises (e.g., nitrogen fixing crops). Today, more powerful tools are being developed to help make biology easier to engineer via an emerging field of research known as “synthetic biology.” Using early versions of these new tools, researchers have begun constructing genomes – the entire DNA program encoding an organism – from scratch². Catalogs containing thousands of standardized DNA parts are being produced and freely distributed³. Undergraduates and high school students are developing genetic programs of their own designs such as bacteria that take living photographs, smell as bananas, detect and warn of arsenic contaminated well water, or provide probiotic supplements⁴.

The Imperative of Synthetic Biology: A Proposed National Research Initiative

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1. Invest in DNA synthesis & construction tech.
2. Invest in open libraries of standard biological parts
3. Explore and test improved legal frameworks for biotech.
4. Develop & implement integrated strategy for biosecurity

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